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<b>(54) Title:</b> LIPOPROTEIN ASSAYS USING ANTIBODIES TO A PAN NATIVE EPITOPE AND RECOMBINANT ANTI-GENS		
<b>(57) Abstract</b>  Methods and compositions are described for determining the level of low density lipoproteins (LDL) in plasma. Native apoprotein B-100 (apo B-100) present in LDL particles is immunologically mimicked by a polypeptide of the invention. A polypeptide includes an amino acid residue sequence corresponding to a pan epitope region of the target apoprotein. A preferred polypeptide is a fusion porotein that simultaneously mimics native apo B-100 and native apo A-I. Improved assay systems and methods for determining HDL and LDL levels in a body fluid sample are also described.		

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LIPOPROTEIN ASSAYS USING ANTIBODIES TO A  
PAN NATIVE EPITOPE AND RECOMBINANT ANTIGENS

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Description

Technical Field

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The present invention relates to assays of lipoprotein markers for abnormal lipid metabolism. The invention particularly relates to assays of human plasma for lipoproteins containing apoproteins B-100 and A-I, and the use of a polypeptide that contains a

15 pan epitope of at least the apo B-100 protein along with antibodies that immunoreact with at least that epitope.

Background

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Lipoproteins are the primary carriers of plasma cholesterol, triacylglycerols, and other lipids. The lipoproteins are micellar lipid-protein complexes consisting of a hydrophobic lipid core surrounded by a shell of polar lipids and apoproteins.

25 Lipoproteins are characterized by their buoyant densities which have resulted in identification of at least six major density classes: chylomicrons, very low-density lipoproteins (VLDL), intermediate-density lipoproteins (IDL), low-density lipoproteins (LDL),

30 lipoprotein(a)s (Lp(a)), and high-density lipoproteins (HDL). At least ten apoproteins associated with these lipoproteins have been isolated and characterized: A-1, A-2, A-4, B-48, B-100, C-1, C-2, C-3, E, and apo(a). All of these apoproteins, except B-48 and

B-100, are water-soluble and exchange readily between lipoproteins of different classes.

Many studies have now established an inverse relationship between plasma HDL cholesterol levels and risk of coronary artery disease (CAD), i.e., elevated levels of plasma cholesterol found in HDL particles correlate with a reduced risk of CAD. See, e.g., Goldbourt et al., Int. J. Epidemiol., 15:51-55 (1986). Similarly, many studies have shown that plasma levels of apoprotein A-I (apo A-I), the major protein component of HDL, are also inversely related to the risk of CAD.

Because of the inverse correlation of HDL levels with CAD, extensive research has been directed towards determining the role of HDL in lipid metabolism. Functionally, HDL is now believed to mediate the removal of cholesterol released into the plasma from dying cells and from membranes undergoing turnover. It is believed that an acyl transferase in HDL esterifies these cholesterol molecules, which are then rapidly shuttled to VLDL or LDL particles by a transfer protein. See, generally, Stryer L., Biochemistry, 3rd ed., W.H. Freeman and Co., New York, (1988), pp 560-564.

The amino acid residue sequence of human apo A-I has been determined by Edman degradation of the protein [Brewer et al., Biochem. Biophys. Res. Comm., 80:623-630 (1978)] and by sequencing full-length apo A-I cDNA [Seilhamer et al., DNA, 3(4):309 (1984)]. These studies report that mature apo A-I is a single chain protein of 243 amino acid residues.

The conformation of the A-I protein in HDL particles is not known [Curtiss et al., J. Biol. Chem., 263:13779 (1988)]. Additionally, immunochemical characterization of native apo A-I,

i.e., apo A-I as it is found on HDL particles, has been problematic because of its antigenic heterogeneity and instability. The antigenic heterogeneity of native apo A-I appears to be due to  
5 masking of some epitopes by lipids in the intact HDL particle or due to a dependency of the antibody-binding ability of some epitopes upon conformations of native apo A-I in the presence of lipids or other HDL-associated proteins, e.g., the A-2 protein. The  
10 antigenic instability of native apo A-I, as manifested by its changing immunoreactivity over time with defined antisera, appears to be due to such phenomena as self-association and deamidation, both of which have been shown to occur in vitro. See Curtiss et al.  
15 in "Proceeding of the Workshop on Lipoprotein Heterogeneity", ed. by Lippel, National Institutes of Health Publication No. 87-2646, pp. 363-377 (1987). Further, the effects of storage and NaOH treatment on native apo A-I immunoreactivity are similar, although  
20 not identical, which suggests that other processes may be involved in the loss of immunoreactivity during storage than deamidation alone [Milthorp et al., Arterio., 6:285-296 (1986)].

The antigenic heterogeneity and instability of  
25 native apo A-I have made it difficult to develop immunoassays for native apo A-I in patient vascular fluid samples. This is due, at least in part, to proposed methods requiring the use of a reference material (standard) that has an immunoreactivity with  
30 anti-apo A-I antibodies comparable to the immunoreactivity of native apo A-I with the anti-apo A-I antibodies.

Recently, efforts to overcome the problems associated with the antigenic heterogeneity and  
35 instability of native apo A-I have focused on using

monoclonal antibodies (Mabs) to identify epitopes on native apo A-I molecules, which expression is consistent or "conserved" under specific isolation and storage conditions. Such epitopes are referred to as  
5 "conserved native epitopes".

Conserved native apo A-I epitopes are reported to be defined by the 1-15 and 90-105 amino acid sequences of apo A-I [Curtiss et al., J. Biol. Chem., 263: 13779 (1988)]. Synthetic polypeptides  
10 representing residues 1-15 and 90-105 are reported to inhibit HDL-binding of monoclonal antibodies AI-16 and AI-18, respectively. The synthetic polypeptide representing apo A-I residue sequence 90-105 and the AI-18 monoclonal antibodies are the subjects of  
15 allowed U.S. Serial No. 07/116,248. Other monoclonal antibodies include Mabs AI-4, AI-7, AI-9, and AI-11, which are reported in U.S. Patent No. 4,677,057 to be immunoreactive with native apo A-I; however, the regions of apo A-I that define the epitopes bound by  
20 these antibodies have not been reported. The disclosures of U.S. Patent No. 5,055,396 and U.S. Patent No. 4,677,057 are incorporated herein by reference.

Additionally, certain monoclonal antibodies  
25 are reported to act as anti-apo A-I "pan" antibodies, i.e., antibodies that bind most, if not all, species of native apo A-I in plasma [Hogle et al., J. Lipid. Res., 29:1221-1229 (1988); Curtiss et al., in "Biotechnology of Dyslipoproteinemias: Clinical  
30 Applications in Diagnosis and Control", Lenfant et al., eds, pp. 217-226, Raven Press (New York), 1989]. Exemplary monoclonal antibodies in this regard include AI-10, AI-11, AI-12, AI-13, and AI-14, as discussed in U.S. Patent No. 5,126,240 which disclosures are  
35 incorporated herein by reference. The epitopes on

native apo A-I molecules with which these antibodies immunoreact have not been identified.

5 An antibody composition that immunoreacts with about 90 percent or more of native apo A-2 is referred to herein as a pan antibody composition, or pan antibodies. That composition can contain polyclonal antibodies, but more preferably contains one or more monoclonal antibodies as are described in the before-cited patents and patent applications. The single  
10 epitope or plurality of epitopes bound by pan antibodies is referred to herein as a pan epitope.

Contrary to the relationship of HDL levels to CAD, high levels of serum LDL have been correlated with abnormal lipid metabolism and CAD [Brown et al.,  
15 New Engl. J. of Med., 323(19): 1289 (1990). During normal lipid metabolism, LDL particles transport cholesterol to peripheral tissues and regulate de novo cholesterol synthesis at these sites. The particles bind to extracellular LDL receptors at these sites and  
20 are taken into the cells by endocytosis [Anderson et al., Cell, 10:351 (1977)]. The particles are chemically broken down within the cell, releasing cholesterol and amino acids. The released cholesterol molecules control feedback mechanisms that regulate de  
25 novo cholesterol synthesis and control synthesis of cholesterol receptors by the cells. See, generally, Stryer L., supra.

The basis for the relationship between LDL and CAD is believed to be due primarily to deposition of  
30 cholesterol on the arterial intima, in the form of atheromatous plaques, when high concentrations of LDL-cholesterol occur in the plasma. High plasma levels of LDL can occur when the diet is enriched in cholesterol or when certain genetic disorders, such as  
35 familial hypercholesterolemia (FH), are present. The

molecular defect in most cases of FH is an absence or deficiency of functional LDL receptors. Homozygotes of FH typically have severely elevated cholesterol levels (680 mg/dl) and usually die of CAD in  
5 childhood. Heterozygotes (1 in 500 persons) typically show elevated cholesterol levels (300 mg/dl) and have increased CAD risk.

Structurally, LDL particles have a cholesterol core comprising about 1500 esterified cholesterol  
10 molecules surrounded by a shell of phospholipids, unesterified cholesterol molecules, and a single molecule of apoprotein B-100 (apo B-100). The hydrophilic shell of an LDL particle facilitates hydration and suspension of the particle in plasma.  
15 Cellular LDL receptors recognize and bind to native apo B-100, i.e., as it appears in LDL particles, thereby extracting LDL particles from the plasma. Hence, the B-100 protein is critical for effective receptor-mediated uptake and clearance of LDL  
20 particles from circulation. Accordingly, investigators have suggested that plasma levels of native apo B-100 actually may be more predictive of CAD risk than plasma LDL cholesterol levels [Sniderman et al., Proc. Natl. Acad. Sci. U.S.A., 77:604-608  
25 (1980); Sniderman et al., Arteriosclerosis, 10(5):665 (1990); Albers et al., Clinics in Laboratory Medicine, 9(1):137 (1989)].

The B-100 protein is also found in VLDL and IDL particles, which are comprised of endogenous  
30 triacylglycerol and cholesterol ester cores, respectively. A fragment of apo B-100, denoted apo B-48, is found in chylomicrons, which transport dietary triacylglycerols and cholesteryl esters. The apo B-48 molecule is reported to correspond to the N-terminal  
35 47 percent (residues 1-2142) of the apo B-100 molecule



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[Innerarity et al., J. Clin. Invest., 80:1794 (1987); Powell et al., Cell, 50:831 (1987); Chen et al., Eur. J. Biochem., 175:111 (1987)].

5 The complete amino acid residue sequence (4563 residues; 514 kDa) of human apo B-100, as determined by cDNA clones of hepatic mRNAs, has been reported [Knott et al., Nature 323: 734 (1986); Knott et al., Nucl. Acids. Resch., 14:7501 (1986)]. The full cDNA sequence for the human apo B-100 gene, consisting of 10 29 exons, also has been reported [Ludwig et al., DNA 6:363 (1987)].

As the level of lipoproteins in circulation is related to the level of associated apoprotein, several schemes for assaying the apoprotein have been 15 proposed. Immunoassays for native apo B-100 have been proposed which utilize specific antibody-containing antisera, including competitive fluid phase and solid phase radioimmunoassay (RIA), enzyme-linked immunosorbent assay (ELISA), and radial 20 immunodiffusion (RID). Problems associated with these immunoassays include reproducibility and the homogeneity and specificity of the antisera used [Currey et al., Clin. Chem., 24:280-286 (1978); Rosseneu et al., Clin. Chem., 28:427-433 (1983); 25 Albers et al. supra].

Several investigators have reported development of monoclonal antibodies against human B-100 apoproteins. The use of anti-apo B-100 monoclonal antibodies for measuring plasma B-100 30 levels has been proposed [Patton et al., Clin. Chem., 29:1898-1903 (1983); Maynard et al., Clin. Chem., 30:1620-1624 (1984); Young et al., Clin. Chem., 32:1484-1490 (1986); Young et al., Arteriosclerosis, 6:178 (1986)]. In addition, a mixture of anti-apo 35 B-100 monoclonal antibodies in a RID assay for plasma

5 B-100 has been proposed [Marconvina et al., Clin. Chim. Acta, 147:117-125 (1985)]. However, these assay techniques suffer from the requirements of lengthy incubations, repeated centrifugations and/or use of radioactive materials.

10 A number of antibody-binding domains of apo B-100 have been mapped with anti-apo B-100 antibodies [Knott et al., Nature 323: 734 (1986); Krul et al., J.Lipid Res. 29:937 (1988); Pease et al., J.Biol.Chem. 265:553 (1990)]. In these studies, fragments of apo B-100 cDNA were cloned into vectors and expressed as  $\beta$ -galactosidase fusion proteins in E. coli. The fusion proteins were probed with various anti-apo B-100 antibodies. The results of these and other studies suggest that anti-apo B-100 monoclonal antibodies often recognize complex epitopes on native apo B-100 which are not recognized on delipidated apo B-100 [Curtiss et al., J. Biol. Chem., 257:15213 (1982)].

20 The antigenic heterogeneity of native apo B-100, as with apo A-I, is well documented. For instance, epitope expression on native apo B-100 molecules has been found to be modulated by: (1) the composition of associated lipids; (2) temperature of the immunoreaction; (3) the degree of isolation of LDL from its native environment; and (4) genetic variety among individuals. Thus, the identification of conserved, pan native epitopes of apo B-100 is important to any generally applicable assay method.

25 An antibody composition that immunoreacts with a conserved, pan native apo B-100 epitope is similarly referred to as a pan antibody composition or pan antibodies.

30 Because of the unique specificity of monoclonal antibodies, their utility in an assay often

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depends upon the affinity of the antibodies for a target antigen. Moreover, although monoclonal antibodies may have sufficient liquid-phase affinity for an antigen, when the same antibodies are affixed to a surface they may not be as effective in binding the antigen. Hence, while certain monoclonal antibodies have been reported, their utility in a particular assay format must be demonstrated. Similarly, when an antigen such as LDL or other apo B-100-containing material is affixed to a surface the affinity of the immobilized material for antibodies in solution may not be sufficient, even though it is sufficient when the apo B-100-containing material is present in the liquid phase. See, e.g., Goding, J., Monoclonal Antibodies: Principles and Practice, Academic Press, New York (1983), pp 40-46.

Currently proposed methods for assaying the lipoproteins described above are deficient in a number of respects. For instance, immunoassay methods for determining the amount of native apo A-I in a lipoprotein-containing sample are disclosed in U.S. Patent No. 4,677,057, U.S. Patent No. 5,126,240 and U.S. Patent No. 5,055,396. However, these methods do not disclose concurrent determination of other apoprotein levels. On the other hand, U.S. Patent No. 4,828,986 proposes competitive and "sandwich" immunoassay methods for determining the ratio of apolipoproteins B-100 and A-I using monoclonal antibodies. However, the latter method does not permit assaying for apolipoproteins B-100 and A-I in the same sample aliquot.

Significantly, previously proposed assay methods require the use of "secondary" serum standards in order to make accurate determinations of the assayed apolipoproteins. As used herein, a "secondary

standard" refers to: (1) delipidated lipoproteins with added native apoprotein; (2) truncated apoproteins; or (3) freshly isolated pooled plasma collected from normal donors, as described in U.S. Patent No.

5 4,828,986. The frequent impracticability of using primary (native) standards is believed to be due to problems associated with their isolation, solubility, stability, and heterogeneity. A review of some of the problems associated with immunoassays of  
10 apolipoproteins is presented in Rosseneu et al., Clin. Chem., 24:280-286 (1978); Albers et al., Clinics in Laboratory Medicine, (9)1:137 (1989).

Since the risk of coronary artery disease is known to be correlated to serum levels of HDL and LDL,  
15 which levels are related to apo A-I and apo B-100, respectively, an assay method that permits determination of both apoproteins is desired. Preferably, the apoprotein assays could be performed on a single sample aliquot. An assay method that  
20 permits ready calibration without the need for secondary serum standards is particularly desired.

#### Summary of the Invention

The present invention relates to a nonnatural  
25 polypeptide that immunologically mimics a pan epitope present in native apo B-100 and in stored samples of that protein. Thus, a polypeptide of the invention immunoreacts with particular antibodies that are also immunoreactive with native apo B-100. Such a  
30 polypeptide is often referred to herein as a "subject polypeptide". A subject polypeptide is dispersible in aqueous media in the substantial absence of denaturing surfactants such as sodium dodecyl sulfate (SDS). Exemplary antibodies include polyclonal antisera to  
35 native apo B-100 and certain "pan" monoclonal

antibodies, such as the antibodies secreted by hybridoma MB47, having ATCC Accession No. HB 8746.

A preferred dispersible polypeptide of the invention contains an amino acid residue sequence of apo B-100, as shown in SEQ ID NO:1 and in Figure 1. Preferably, the apo B-100 sequence comprises about 80 up to about 375 residues as shown in the figure that includes amino acid residues about 3430 through about residue 3510, corresponding to residues 217 through 297 of SEQ ID NO:1. More preferably, the polypeptide contains an amino acid residue sequence of apo B-100 of about 90 through about 160 residues in length that includes residues about 3430 through about residue 3520, corresponding to residues 217 through 307 of SEQ ID NO:1. A particularly preferred subject polypeptide is a recombinant polypeptide that is expressed fused at the carboxy-terminus of another polypeptide or protein sequence.

In another aspect of the invention, a bifunctional apo A-I/B-100 fusion polypeptide that is dispersible in the substantial absence of denaturing surfactants is contemplated. The fusion polypeptide immunologically mimics both the before-described pan native apo B-100 epitope and a pan native apo A-I epitope. Thus, the fusion polypeptide can be regarded as a recombinant antigen that immunoreacts with antisera to a pan native apo B-100 epitope and antisera to a pan native apo A-I epitope, including monoclonal antibodies to these apoproteins.

Structurally, a dispersible apo A-I/B-100 fusion polypeptide of the invention contains:

(a) a first amino acid residue sequence of apo A-I as shown in SEQ ID NO:3 and in Figure 2 that includes at least the amino acid sequence positions 120 through 135, with the fusion polypeptide also

immunoreacting with pan anti-apo A-I antibodies, such as those secreted by a hybridoma selected from the group consisting of: AI-4 (ATCC HB 8744), AI-7 (ATCC HB 8745), AI-9 (ATCC HB 8741), AI-10 (ATCC HB 9200),  
5 AI-11 (ATCC HB 9201), AI-12 (ATCC HB 9202), AI-13 (ATCC HB 9203), AI-14 (ATCC HB 9204), and AI-18 (ATCC HB 9570); and

(b) a second amino acid residue sequence of apo B-100 as shown in Figure 1 from position about  
10 3430 through about 3510, corresponding to residues 217 through 297 of SEQ ID NO:1, with the fusion polypeptide immunoreacting with pan anti-apo B-100 antibodies, such as those secreted by the hybridoma MB47 (ATCC HB 8746). The second amino acid residue  
15 sequence is operatively linked to the first amino acid residue sequence and constitutes the carboxy-terminal portion of the fusion polypeptide.

An apo A-I/B-100 fusion polypeptide contains an apo B-100 amino acid residue sequence described  
20 before for a subject polypeptide and an apo A-I amino acid residue sequence of about 15 up through about 250 residues in length. Also, the fusion polypeptide can have the apo A-I amino acid residue sequence linked to the apo B-100 sequence via a linking amino acid  
25 residue sequence containing one or more residues.

A subject apo A-I/B-100 fusion polypeptide thus contains a first amino acid residue sequence of apo A-I from about residue 120 through about residue 135 operatively linked to a second amino acid residue  
30 sequence that includes an apo B-100 sequence from about amino acid residue 3430 through about residue 3510. The former apo A-I sequence can be represented by the formula:

LysValGlnProTyrLeuAspAspPheGlnLysLysTrpGlnGluGlu  
 1                      5                      10                      15  
 (SEQ ID NO:5),

5 as described more fully hereinafter. More preferably, an apo A-I/B-100 fusion polypeptide contains the amino acid residue sequence of apo B-100 from about residue 3430 through about residue 3520 operatively linked to the amino acid residue sequence of apo A-I from about residue 19 through about residue 250 of SEQ ID NO:3.

10 Most preferably, a subject fusion polypeptide has a beta-galactosidase moiety fused to the N-terminus of the polypeptide as an artifact of the cloning vector used for expression of the polypeptide. Thus, from N-terminus to C-terminus, a most preferred fusion protein contains the following order of amino acid residue sequences: beta-galactosidase apo A-I/apo B-100.

20 Also contemplated is an isolated DNA segment comprising a nucleotide sequence (SEQ ID NO:2) encoding an amino acid residue sequence of apo B-100 as shown in Figure 1 including the sequence from about residue 3430 through about residue 3510, corresponding to residues 219 through 297 of SEQ ID NO:1 and nucleotides 649 through 891 of SEQ ID NO:2. The DNA segment preferably contains about 240 through about 1200, and more preferably about 295 through about 485, base pairs that encode an apo B-100 amino acid sequence.

30 Further contemplated is a self-replicating vector (recombinant DNA molecule) that comprises a before-described DNA segment encoding an amino acid residue sequence of apo B-100 as shown in Figure 1 from about residue 3430 through about residue 3510. Preferably, a vector of the invention is transformed and expressed in the SURE strain of E. coli. An

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exemplary vector in this regard encodes a beta-galactosidase amino acid residue sequence operatively linked to the apo B-100 sequence, as in a fusion protein.

5           A DNA segment encoding both a before-described apo B-100 polypeptide and an apo A-I polypeptide and an expression vector containing that DNA segment operatively ligated therein for expression is also contemplated as a recombinant DNA molecule. A  
10 particularly preferred DNA segment encodes a fusion polypeptide that includes an N-terminal beta-galactosidase, an apo A-I polypeptide described before and a C-terminal apo B-100 subject polypeptide.

15           A method for assaying a body fluid sample for its native apo B-100 content is also contemplated. Thus, one embodiment comprises a solid phase method for assaying the amount of native apo B-100 in a body fluid sample by competitive binding.

20           Here, in step (a) a predetermined amount of body fluid sample containing native apo B-100 is admixed in a liquid medium with a predetermined amount of a before-described dispersible subject apo B-100 polypeptide affixed to a solid support and a predetermined amount of pan anti-apo B-100 antibodies  
25 to form a solid-liquid admixture. The pan anti-apo B-100 antibodies immunoreact with the native apo B-100 of the sample and with the polypeptide.

30           The solid-liquid admixture is maintained under biological assay conditions for a time period sufficient for the pan anti-apo B-100 antibodies to immunoreact with the polypeptide to form a solid-phase polypeptide/anti-apo B-100 immunocomplex and with the native apo B-100 in the liquid phase to form an apo B-100/anti-apo B-100 immunocomplex in the liquid  
35 phase.



The amount of solid-phase polypeptide/anti-apo B-100 immunocomplex formed is determined, and thereby the amount of native apo B-100 in the body fluid sample is also determined.

5 Similarly contemplated is a method for assaying the amounts of native apo B-100 and native apo A-I in a body fluid sample. This method comprises the following steps.

10 (a) A predetermined amount of body fluid sample is mixed in a liquid medium with predetermined amounts of (i) a dispersible subject apo A-I/B-100 fusion polypeptide affixed to a solid support, (ii) pan anti-apo B-100 antibodies, and (iii) pan anti-apo A- antibodies to form a solid-liquid admixture. The  
15 anti-apo B-100 antibodies immunoreact with the native apo B-100 of the sample and the fusion polypeptide, and the anti-apo A-I antibodies immunoreact with the native apo A-I of the sample and the fusion polypeptide.

20 (b) The solid-liquid admixture is maintained under biological assay conditions for a time period sufficient for the anti-apo B-100 antibodies and anti-apo A-I antibodies to immunoreact with the fusion polypeptide and the liquid phase apo B-100 and apo A-I  
25 proteins to form solid phase-bound fusion polypeptide/anti-apo B-100 and fusion polypeptide/anti-apo A-I immunocomplexes and liquid phase apo B-100/anti-apo B-100 and apo A-I/anti-A-I immunocomplexes.

30 (c) The amounts of solid-phase fusion polypeptide/anti-apo B-100 and fusion polypeptide/anti-apo A-I immunocomplexes formed are determined, and thereby the amounts of native apo B-100 and native apo A-I in the sample are also  
35 determined.

Also contemplated is a method for assaying the amount of native apo B-100 in a body fluid sample that employs a subject polypeptide to inhibit the reaction of native apo B-100 with anti-apo B-100 antibodies.

5 Here, the method comprises the following steps.

(a) A predetermined amount of body fluid sample containing native apo B-100 is admixed in a liquid medium with predetermined amounts of a dispersible subject polypeptide and solid phase-bound  
10 pan anti-apo B-100 antibodies to form an admixture, with the anti-apo B-100 antibodies immunoreacting with the native apo B-100 of the sample and with the polypeptide.

(b) The admixture is maintained under  
15 biological assay conditions for a time period sufficient for the anti-apo B-100 antibodies to immunoreact with the polypeptide and native apo B-100 to form a polypeptide/anti-apo B-100 immunocomplex and an apo B-100/anti-apo B-100 immunocomplex.

(c) The amount of polypeptide/anti-apo B-100  
20 immunocomplex formed is determined, and thereby the amount of native apo B-100 in the body fluid sample is also determined.

Further contemplated is a method for  
25 standardizing an assay for the amount of native apo B-100 in a body fluid sample comprising:

(a) admixing a predetermined amount of body fluid sample containing native apo B-100 in a liquid medium with a predetermined amount of pan anti-apo  
30 B-100 antibodies to form a first admixture, the anti-apo B-100 antibodies immunoreactive with the native apo B-100 of the sample and with a before-described, subject apo B-100 polypeptide;

(b) maintaining the first admixture under  
35 biological assay conditions for a time period

sufficient for the pan anti-apo B-100 antibodies to immunoreact with the native apo B-100 in the sample to form a native apo B-100/anti-apo B-100 immunocomplex;

5 (c) admixing a predetermined amount of the pan anti-apo B-100 antibodies in a liquid medium with a predetermined amount of a before-described, dispersible subject apo B-100 polypeptide to form a second admixture;

10 (d) maintaining the second admixture under biological assay conditions for a time period sufficient for the anti-apo B-100 antibodies to immunoreact with the polypeptide to form a polypeptide/anti-apo B-100 immunocomplex; and

15 (e) determining the amount of native apo B-100/anti-apo B-100 immunocomplex formed and the amount of polypeptide/anti-apo B-100 immunocomplex formed, and thereby the amount of native apo B-100 in the body fluid sample.

20 Similarly contemplated is a method for standardizing an assay for the amounts of native apo B-100 and native apo A-I in a body fluid, as described hereinafter.

The present invention thus affords improved compositions and methods for assaying native apo B-100  
25 levels, that are related to LDL cholesterol levels, in a body fluid sample. The present invention also affords improved compositions and methods for assaying native apo B-100 levels in conjunction with native apo A-I levels, the level of apo A-I being related to the  
30 HDL cholesterol levels. The immunoassays contemplated within the present invention can be performed more expeditiously and accurately than by current methods.

### Brief Description of the Drawings

Figure 1 shown in four sheets (Figs. 1A, 1B, 1C and 1D) depicts the amino acid residue sequence of human apo B-100, and its corresponding cDNA, from amino acid residue 3214 through residue 3590 according to the numbering scheme of Ludwig et al., DNA, 6:363 (1987). The amino acid residue sequence is also referred to herein as SEQ ID NO:1, and the DNA sequence is referred to herein as SEQ ID NO:2.

Figure 2 shown in three sheets as Figs. 2A, 2B and 2C illustrates the amino acid residue sequence of human apo A-I, including presignal (residues 1-18) and propeptide (residues 19-24) sequences, according to Seilhamer et al., DNA, 3(4):309 (1984). The corresponding "sense" cDNA strand for residues 1-267 is also presented. The amino acid residue sequence is also referred to herein as SEQ ID NO:3, with the DNA sequence being referred to as SEQ ID NO:4.

### Detailed Description of the Invention

#### A. Definitions

Amino Acid Residue Sequence: a series of two or more amino acid residues joined via peptide linkages between adjacent residues to form a peptide or polypeptide. An amino acid residue sequence is conveniently represented by the one or three letter abbreviations for its constituent amino acids. The abbreviations used herein for amino acids are those provided at 37 C.F.R. §1.822(b)(2) and are reproduced in the following table of correspondence:

		<u>ABBREVIATION</u>		<u>AMINO ACID</u>
		<u>1-Letter</u>	<u>3-Letter</u>	
		Y	Tyr	tyrosine
		G	Gly	glycine
5		F	Phe	phenylalanine
		M	Met	methionine
		A	Ala	alanine
		S	Ser	serine
		I	Ile	isoleucine
10		L	Leu	leucine
		T	Thr	threonine
		V	Val	valine
		P	Pro	proline
		K	Lys	lysine
15		H	His	histidine
		Q	Gln	glutamine
		E	Glu	glutamic acid
		Z	Glx	Glu and/or Gln
		W	Trp	tryptophan
20		R	Arg	arginine
		D	Asp	aspartic acid
		N	Asn	asparagine
		B	Asx	Asn and/or Asp
		C	Cys	cysteine
25		J	Xaa	Unspecified

The individual residues comprising an amino acid  
 residue sequence herein may be in the D or L isomeric  
 form as long as the desired functional property is  
 30 retained by molecule(s) incorporating the amino acid  
 residue sequence. Also, the amino acid residue  
 sequence may include post-translationally modified  
 amino acids, e.g., hydroxylated, glycosylated amino  
 acid residues, or residues linked via disulfide bonds.  
 35 In addition, an amino acid residue sequence can

include one or more modified or unusual amino acids, such as those listed in 37 C.F.R. §1.822(b)(4), which are incorporated herein by reference. An amino acid residue sequence can be represented by the

5 abbreviations corresponding to its constituent amino acids in which a hyphen between two adjacent abbreviations indicates a peptide linkage between the corresponding residues.

Antibody: a polypeptide that immunologically

10 binds to a ligand group. Antibodies, as used herein, are immunoglobulin molecules and immunologically active fragments of immunoglobulin molecules. Such portions known in the art as Fab, Fab'; F(ab')<sub>2</sub> and F<sub>v</sub> are included. Typically, antibodies bind ligands that

15 range in size from about 6 through about 34 Å with association constants in the range of about 10<sup>4</sup> to 10<sup>10</sup> M<sup>-1</sup>, and as high as 10<sup>13</sup> M<sup>-1</sup>. Antibodies can bind a wide range of ligands, including small molecules such as steroids and prostaglandins, biopolymers such as

20 nucleic acids, proteins and polysaccharides, and synthetic polymers such as polypropylene. An "antibody combining site" is that structural portion of an antibody molecule comprised of a heavy and light chain variable and hypervariable regions that

25 specifically binds (immunoreacts with) antigen. The term "immunoreact" in its various forms is used herein to refer to binding between an antigenic determinant-containing molecule (antigen) and a molecule containing an antibody combining site such as a whole

30 antibody molecule or a portion thereof. An "antigenic determinant" is the structural portion of the antigen that is immunologically bound by an antibody combining site. The term is also used interchangeably with "epitope". Antibodies can bind a single epitope of an

antigen (monoclonal) or multiple epitopes (polyclonal).

5                    Fusion protein or polypeptide: A protein or polypeptide containing amino acid residue sequences from each of two distinct proteins; it is formed by the expression of a recombinant gene in which two coding sequences have been joined together such that their reading frames are in phase. A fusion protein results when the expressed material contains at least  
10                   one complete protein sequence, whereas only portions of the proteins are present in a fusion polypeptide. Hybrid genes of this type may be constructed in vitro, e.g. in order to label the product of a particular gene with a protein which can be more readily assayed.  
15                   For example, if a gene is fused to lacZ in Escherichia coli, a fusion protein or polypeptide with  $\beta$ -galactosidase activity or a partial sequence may be obtained. Alternatively, a protein may be linked to a signal peptide to allow its secretion by the cell.

20                   Ligand: a molecule having a structural region that binds specifically to a particular receptor molecule, usually via electrostatic forces and/or hydrogen bonds. An exemplary receptor molecule is an antibody combining site.

25                   Oligonucleotide/Polynucleotide/DNA Segment: a polymer of single or double stranded nucleotides. As used herein, these terms and their grammatical equivalents, can be composed of a naturally-occurring nucleic acid, e.g., adenine (A), cytosine (C), guanine (G), thymine (T), uracil (U), inosine (I), as well as  
30                   a modified base, as presented at 37 C.F.R. §1.82(p)(1).

Peptide/Polypeptide: a polymer comprising at least two amino acid residues in which adjacent  
35                   residues are connected by a peptide bond between the

alpha-amino group of one residue and the alpha-carbonyl group of an adjacent residue. The primary structure of a polypeptide has a primary amine group at one terminus and a carboxylic acid group at the other terminus of the polymer. Thus, a polypeptide may be represented by the formula:



where R is a side chain characteristic of a given amino acid residue and i indicates the number of amino acid residues comprising the polymer which number is two or more. A polypeptide can comprise one or more amino acid residue sequences. Also, a polypeptide in aqueous solution is usually in one or more zwitterionic forms depending on the pH of the solution.

Protein: a single polypeptide or set of cross-linked polypeptides comprising more than about 50 amino acid residues. Proteins can have chemical crosslinking, e.g., via disulfide bridges, within the same polypeptide chain or between adjacent polypeptides. When a protein is glycosylated it may be called a glycoprotein. When a protein comprises one or more discrete polypeptide/protein subunits linked together, as by a peptide linkage, amino acid residue sequence, disulfide bridge, and the like, the protein is frequently termed a fusion protein, fusion polypeptide, chimeric fusion, and the like.

Receptor: a biologically active proteinaceous molecule having a structural region that specifically binds to (or with) another molecule (ligand). An antibody combining site is one type of receptor.

Vector: a DNA molecule capable of autonomous replication in a cell and to which a DNA segment,



e.g., gene or polynucleotide, can be operatively linked so as to bring about replication of the attached segment. An expression vector is capable of directing expression of a DNA segment encoding one or more polypeptides such as when the DNA segment is under the control of a promoter sequence of the vector.

10                   B.    Polypeptides that Immunologically Mimic apo B-100

                  The present invention relates to an isolated polypeptide that immunologically mimics a pan immunologic binding region (epitope) of native apo B-100; i.e., the polypeptide immunoreacts with antibodies that immunoreact with at least 90 percent and more preferably at least 95 percent of radioiodinated apo B-100. A subject polypeptide mimics a single pan epitope of apo B-100. Moreover, a subject polypeptide can immunologically mimic one or more other apolipoprotein epitopes, such as a pan native apo A-I epitope, in addition to mimicking a pan native apo B-100 epitope.

                  Thus, a polypeptide is contemplated that mimics a plurality of apolipoprotein epitopes, including an apo B-100 epitope. When a polypeptide of the invention immunologically mimics more than one class of apolipoprotein, including apo B-100, the polypeptide at least immunoreacts with particular pan anti-apo B-100 antibodies, and also immunoreacts with antibodies immunoreactive with the other apolipoprotein.

                  A preferred polypeptide of the invention immunoreacts with pan anti-apo B-100 antibodies secreted by the hybridoma designated MB47, having ATCC Accession No. HB 8746. The monoclonal antibodies

secreted by hybridoma MB47 are referred to herein as monoclonal MB47 antibodies or MB47 antibodies. MB47 antibodies immunoreact substantially identically with both with freshly drawn apo B-100 and with stored samples that have partially decomposed. Thus, a subject polypeptide can be used as a long sought absolute immunological standard for apo B-100.

An exemplary polypeptide of the invention that immunoreacts with antibodies immunoreactive with more than one class of apolipoprotein is a fusion polypeptide immunoreactive with pan anti-apo B-100 antibodies and pan anti-apo A-I antibodies. Preferably, such a fusion polypeptide immunoreacts with MB47 antibodies as well as with pan anti-apo A-I antibodies secreted by the hybridoma designated AI-11, having ATCC Accession No. HB 9201.

A subject polypeptide that immunologically mimics native apo B-100 typically comprises an amino acid residue sequence identical to, or conservatively substituted from (as described hereinafter), a human apo B-100 amino acid residue sequence. Thus, a subject polypeptide can comprise a "truncated" amino acid residue sequence of apo B-100 up through about 375 amino acid residues in length, and includes an amino acid residue sequence that defines a pan antibody-binding region (epitope) of native apo B-100. A subject polypeptide can also include an immunologically "inert" spacing (linking) amino acid residue sequence, such as a  $\beta$ -galactosidase sequence (MW =  $1.1 \times 10^5$  Da; about 1200 residues), as long as the immunologically inert sequence does not substantially interfere with the immunological binding properties of the polypeptide; i.e., as immunologically inert, as described herein.

5 A subject polypeptide is dispersible (soluble)  
in an aqueous medium such as a usual buffer like PBS  
or bicarbonate in the substantial absence of a  
denaturing surfactant such as sodium dodecyl sulfate  
(SDS), polyoxyethylene(9)octylphenyl ether (Nonidet  
P-40 or Triton X-100), or the like. Small, non-  
denaturing amounts of such surfactants that do not  
interfere with binding of a fusion polypeptide to the  
surface of a plastic microtiter plate well can be  
10 present. Small molecule non-detergent denaturants  
such as urea, guanidine and 2-mercaptoethanol can be  
present in the dispersions.

As such, a subject polypeptide exemplifies an  
unexpected advantage over seemingly similar  
15 polypeptides known in the art such as those described  
in Pease et al., J. Biol. Chem., 265:553-568 (1990),  
all of which are insoluble and require the presence of  
SDS even for use in Western blots. The required SDS  
dispersibility of those known polypeptides prevents  
20 their use in solid phase assays using plastic  
microtiter plates because the SDS inhibits binding of  
the polypeptide to the plastic solid phase microtiter  
plate well. SDS-containing aqueous dispersions of  
such insoluble materials are also turbid, viscous and  
25 cannot readily be pipetted. A subject polypeptide is  
therefore often referred to herein as dispersible or  
as a dispersible subject polypeptide.

A preferred polypeptide contains 80 residues  
through about 375 residues and includes the amino acid  
30 residue sequence of apo B-100 from about residue 3430  
through about residue 3510 as shown in Figure 1  
corresponding to residues 217 through 297 of SEQ ID  
NO:1. A more preferred polypeptide contains 90 or  
more residues and includes an apo B-100 sequence from  
35 about position 3430 through about position 3520

corresponding to residues 217 through 307 of SEQ ID NO:1. Other preferred polypeptides include an amino acid residue sequence of apo B-100 as exemplified by sequences s1-s9 in Table 1 below.

5

TABLE 1  
Selected Truncated Amino Acid and  
cDNA Sequences of apo B-100<sup>1</sup>

No.	N-terminus <sup>2</sup>		C-terminus <sup>2</sup>		Sequence <sup>3</sup> Length	Relative MB47 Binding <sup>4</sup>
	A.A.#	Nucl.#	A.A.#	Nucl.#		
s1	3429	10287	3510	10530	81	++
s2	3418	10254	3510	10530	92	++
s3	3386	10158	3510	10530	124	++
s4	3353	10059	3510	10530	157	++
s5	3429	10287	3523	10569	94	+++
s6	3429	10287	3544	10632	115	+++
s7	3429	10287	3565	10695	136	+++
s8	3429	10287	3590	10770	161	+++
s9	3214	9642	3590	10770	376	+++

<sup>1</sup>Sequence numbering scheme according to Ludwig et al. DNA, 6:363 (1987).

<sup>2</sup>A.A.# and Nucl.# are amino acid and nucleotide positions, respectively.

<sup>3</sup>The number of amino acid residues in the sequence is indicated.

<sup>4</sup>MB47 binding studies are described in Example 1.

The length and apo B-100 sequence positions of a subject polypeptide are of importance to the invention, as is discussed below.

An overlapping series of synthetic polypeptides, each containing 20 to 30 residues was prepared. The sequences of those polypeptides corresponded to the apo B-100 sequence from amino acid residue position 3416 through 3505. No binding was

observed between any of the synthetic polypeptides and monoclonal MB47.

A further series of four expression polypeptides was also prepared each of which contained  
5 15, 30, 37 or 53 amino acid residues and whose sequences corresponded to positions ending at position 3510 of the B-100 sequence. Assays using monoclonal MB47 also exhibited negative binding results.

Thus, even through there can be no disulfide  
10 bridges and no turn-inducing prolines in the region studied, the complete apo B-100 pan epitope bound by monoclonal MB47 appears to be non-linear and due to a folded, secondary and/or tertiary structure of the apo B-100 protein and a subject polypeptide. That epitope  
15 is mimicable by a polypeptide having an apo B-100 sequence that contains about 80 amino acid residues up through about 375 residues, more preferably up through about 160 residues, and includes an apo B-100 amino acid residue sequence from amino acid residue position  
20 about 3430 (about nucleotide position 10290) through about residue 3510 (about nucleotide position 10560). Polypeptides s1-s9 of Table 1 are exemplary of these polypeptides.

A more preferred polypeptide, also useful for  
25 assays or as a standard, is also present in a polypeptide containing about 90 through about 375 amino acid residues, and more preferably up through about 160 residues of the apo B-100 amino acid residue sequence from about residue position 3430 (about  
30 nucleotide position 10,290) through about residue position 3520 (about nucleotide position 10,570). This more preferred group includes polypeptides s5-s9 of Table 1.

A schematic that illustrates the relative  
35 lengths and positions of useful polypeptides s1-s8, as

well as shorter recombinant apo B-100 amino acid sequence-containing polypeptides is provided below in Table 2.

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TABLE 2

Schematic of Apo B-100 Sequence-  
Containing Polypeptides

<u>Polypeptide Nucleotide Sequence Position</u>	<u>Polypeptide Designation of Table I</u>	<u>Relative MB47 Immuno- reactivity**</u>
(10,059) 471 bp (10,530) I-----I	S4	++
(10,158 372 bp I-----I	S3	++
(10,254) 276 bp I-----I	S2	++
(10,287) 243 bp I-----I	S1	++
(10,359) 171 bp I-----I		-
(10,419) 111 bp I-----I		-
(10,440) 90 bp I-----I		-
(10,485) 45 bp I-----I		-
(10,287) 483 bp (10,770) I-----I	S8	+++
408 bp (10,695) I-----I	S7	+++
345 bp (10,632) I-----I	S6	+++
282 bp (10,569) I-----I	S5	+++

\* Parenthesized numbers are 5' and 3' positions in the apo B-100 gene shown in Figure 1.

\*\* Strong binding = +++; good binding = ++; no binding = -.

The results obtained herein are contrary to those reported in Table 1 of Pease et al., J. Biol. Chem., 265:553-568 (1990). It should first be noted that those authors' results were obtained using  
5 Western blots with expressed polypeptides dispersible only in the presence of both 2-mercaptoethanol (2-ME) and sodium dodecyl sulfate (SDS).

Second, those authors identified the monoclonal MB47 epitope position at about residue  
10 3506, whereas results with other monoclonals were reported to be in sequences having a length of about 75 through about 600 residues. An earlier report by the same research group [Knott et al., Nature, 323:734 (1986)] had placed the MB47 epitope near the carboxy-  
15 terminus of a recombinant  $\beta$ -galactosidase-apo-B-100 polypeptide containing amino acid residues 3350-3506. The present results with the 20-30-mer overlapping synthetic polypeptides discussed above, and particularly with a polypeptide spanning residues  
20 3491-3510, as well as the results using the recombinants terminating at position 3510 (nucleotide position 10,530) indicate that monoclonal MB47 does not bind to a linear epitope at residue position about 3506. Rather, pan monoclonal MB47 binds to a  
25 conformational epitope that requires residues in the region of residue position 3430, as well as those in the region of positions 3510-3520.

Those earlier workers also found maximal binding to polypeptides containing about 400 residues  
30 at residue positions 3214-3611 and about 510 residues at positions 3214-3728, and poorer binding with a sequence of about 155 residues at residue positions 3351-3506. That latter region and the region of a recombinant prepared herein encompassing amino acid  
35 residue positions 3349 through 3506 substantially



encompass the sequence of polypeptide s5 (positions 3429-3523), which was found to exhibit maximal binding herein.

5       The present results therefore provide a  
surprising finding as to the length and positional  
requirements of the pan apo B-100 epitope bound by  
monoclonal MB47. Thus, a sequence of about 400  
residues or more encompassing the B-100 sequence from  
10       residue position about 3430 through about 3520 plus  
added B-100 amino- and carboxyl-terminal residues can  
provide maximal MB47 binding in a Western blot and in  
the presence of SDS, as does a sequence of about 90  
through about 375 residues in a solid phase assay in  
the substantial absence of SDS, whereas a polypeptide  
15       of about 155 residues whose sequence was displaced  
only about 15 residues toward the amino-terminus from  
residue about 3520 provided poorer, unacceptable  
binding in a Western blot. Still further, Pease et  
al. found a polypeptide of about 360 residues whose  
20       carboxy-terminus was about 25 residues toward the  
amino-terminus from residue position 3420 showed  
minimal to no Western blot binding (binding reported  
as  $\pm$  as compared to  $++$  for the 400-mer above) with  
monoclonal MB47.

25       Preferably, a subject polypeptide is  
recombinantly expressed in the SURE strain of  
Escherichia coli. Thus, a preferred subject  
polypeptide, which immunologically mimics only the pan  
native apo B-100 apolipoprotein epitope, is expressed  
30       by SURE E. coli transformed with a vector encoding the  
polypeptide. A preferred polypeptide in this regard  
is expressed by SURE E. coli transformed with the  
plasmid designated "84", which plasmid encodes  
polypeptide s4 of Table 1. A more preferred  
35       polypeptide is also expressed by SURE E. coli

transformed with the plasmid designated "144", which plasmid encodes polypeptide s5 of Table 1. These and other plasmids are discussed in greater detail hereinafter.

5           As mentioned before, another aspect of the present invention is a dispersible fusion polypeptide, or a fusion protein, that immunologically mimics native apo B-100 and native apo A-I. The entire apo A-I amino acid residue sequence can be operatively  
10 linked to a before-described apo B-100 sequence, making the linked construct a fusion protein, or an apo A-I polypeptide sequence of less than the entire apo A-I length can be used, making the operatively linked construct a fusion polypeptide. Thus, such a  
15 fusion polypeptide or protein immunoreacts with pan anti-apo B-100 antibodies as well as with pan anti-apo A-I antibodies. For convenience of expression, a fusion protein will often hereinafter be referred to as a fusion polypeptide.

20           As is discussed elsewhere, a recombinant polypeptide that immunologically mimics only the pan apo B-100 epitope can also be expressed operatively linked to all or part of the amino acid sequence of another immunologically inert protein, and therefore  
25 also be a fusion polypeptide or fusion protein. To distinguish constructs containing only an apo B-100 epitope from those containing both apo B-100 and apo A-I epitopes, as the relevant epitopes, the former will hereinafter be generally referred to as an apo  
30 B-100 fusion polypeptide, and the latter as an apo A-I/B-100 fusion polypeptide.

          A contemplated fusion polypeptide immunoreacts with anti-B-100 MB47 monoclonal antibodies. Still  
35 more preferred is a fusion polypeptide that includes the apo A-I amino acid residue sequence from position

about 120 through about 135 (the pan epitope) and also immunoreacts with pan anti-apo A-I monoclonal antibodies secreted by hybridoma AI-11 (ATCC HB 9201). An apo A-I-containing fusion polypeptide can also preferably immunoreact with monoclonal antibodies secreted by a hybridoma selected from the group consisting of:

		<u>Deposit Date</u>
	AI-4 (ATCC HB 8744),	3/5/85
10	AI-7 (ATCC HB 8745),	3/5/85
	AI-9 (ATCC HB 8741),	3/5/85
	AI-10 (ATCC HB 9200),	9/16/86
	AI-12 (ATCC HB 9202),	9/16/86
	AI-13 (ATCC HB 9203),	9/16/86
15	AI-14 (ATCC HB 9204),	9/16/84
	AI-18 (ATCC HB 9570),	10/14/87

which hybridomas along with MB47 (deposited 3/6/85), AI-11 (deposited 9/16/86) and MB24 (deposited 3/6/85) have been deposited with the ATCC pursuant to the Budapest Treaty.

A pan native epitope of either apo B-100 or apo A-I can thus be defined by immunoreaction with an appropriate monoclonal antibody such as MB47 or AI-11, respectively. A pan anti-apo B-100 antibody or a pan anti-apo A-I antibody immunoreacts with at least 90 percent, and preferably at least 95 percent of <sup>125</sup>I-labeled apo B-100 or apo A-I, respectively.

A subject polypeptide referred to herein as an apo A-I/B-100 fusion polypeptide is dispersible as discussed previously, and comprises a first amino acid residue sequence that includes an amino acid residue sequence of a pan apo A-I epitope operatively linked to a second amino acid residue sequence that includes an amino acid residue sequence of a pan apo B-100

epitope. An amino acid residue sequence is said to be "operatively linked" to a second amino acid residue sequence when the sequences are joined by one or more covalent bonds, as by a direct peptide bond, disulfide bond, or by an intervening peptide-bonded linking amino acid residue sequence. Preferably, the first and second amino acid residue sequences are linked together via a peptide-bonded third amino acid residue sequence of one or more residues.

10 A fusion polypeptide of the invention preferably includes a pan apo B-100 amino acid residue sequence as listed in Table 1 above. Thus, a preferred apo A-I/B-100 fusion polypeptide includes a truncated apo B-100 sequence listed in Table 1  
15 operatively linked at the carboxy-terminus of an amino acid residue sequence of apo A-I that defines a pan epitope of that molecule; i.e., residues 120 through 135. A particularly preferred apo A-I/B-100 fusion polypeptide includes an amino acid residue sequence of  
20 apo A-I as shown in SEQ ID NO:3 and Figure 2. The apo A-I sequence is according to Seilhamer et al., DNA, 3(4):309 (1984).

A fusion polypeptide of the invention is sufficiently long to permit the polypeptide to display  
25 the immunologic activities described before. Hence, the length of a particular polypeptide of the invention depends upon the immunologic activities desired. The pan apo B-100 and apo A-I epitopes of a fusion polypeptide are arrayed such that  
30 immunoreaction of pan anti-apo B-100 antibodies is substantially independent of immunoreaction of pan anti-apo A-I antibodies.

As a preferred fusion polypeptide is an apo A-1/B-100 polypeptide comprising an amino acid residue  
35 sequence of a pan apo B-100 epitope operatively linked

A more preferred apo A-I/B-100 fusion polypeptide includes an amino acid residue sequence of apo B-100 from about residue 3430 through about residue 3520 (e.g., s5 in Table 1) operatively linked via a linking amino acid residue sequence to an amino acid residue sequence of apo A-I as shown in Figure 2 from about residue 120 through about residue 135 therein. That latter apo A-I sequence is represented by the amino acid residue sequence:

A particularly preferred apo A-I/B-100 fusion polypeptide of the invention includes the amino acid residue sequence from residue 3429 (about 3030) to residue 3523 (about 3520; s5 in Table 1) of apo B-100 linked via a peptide bond to the C-terminal end of the apo A-I amino acid residue sequence from residue 19 through residue 240. Thus, the apo A-I sequence from residue 121 through residue 240 can also be viewed as an about 120 amino acid residue linking sequence. Such a fusion polypeptide is synthesized by the

plasmid designated "137" when transformed and expressed in SURE E. coli, details of which are described hereinafter.

5 Exemplary linking sequences are formed by expression of nucleotide sequences that are utilized for restriction endonuclease cleavage of DNA molecules used to construct the recombinant molecules that express an apo B-100 fusion polypeptide or apo A-1/B-100 fusion polypeptide or protein. Blunted  
10 restriction sites can also give rise to linking amino acid residue sequences upon ligation into a recombinant DNA molecule (as discussed hereinafter) and expression. Amino acid residue sequences of apo B-100 and apo A-I in addition to those of a pan  
15 epitope can also be included as a linking sequence. Another exemplary linking sequence is GlyGlyGlyGlySer (SEQ ID NO:6), whose encoding DNA can be ligated between an apo A-I- and a B-100-encoding DNA sequence.

It should be apparent to any worker skilled in  
20 molecular biology that substantially any polypeptide can be used as a linking sequence so long as it is immunologically inert, as described herein, and so long as it is free of residues such as cysteines and residues such as tryptophans, phenylalanines and the  
25 like that can cause insolubilization (non-dispersibility in the substantial absence of SDS) of the expressed material or otherwise inhibit its immunoreactivity. A linking sequence, also sometimes referred to herein as a spacing sequence, can contain  
30 one residue up through about 1200 residues as is the case when a fusion polypeptide or fusion protein is expressed linked to the carboxy-terminus of  $\beta$ -galactosidase.

The apo B-100 sequence of an instant fusion  
35 polypeptide is located proximate the carboxy-terminus

of the molecule so that the N-terminus of the apo B-100 sequence is linked to the C-terminus of the other, non-apo B-100 sequence such as  $\beta$ -galactosidase. This preference stems from diminished monoclonal MB47 binding that was observed when the order of two sequences in the expressed fusion polypeptide were reversed. Thus, when an apo B-100 epitope is placed at the carboxy-terminus binding as noted in Table 1 is observed, whereas when at the amino-terminus of an apo A-I epitope, binding was questionable (+++ or ++ vs  $\pm$ ).

Accordingly, an apo A-I/B-100 fusion polypeptide has an apo A-I amino acid residue sequence located proximate the amino-terminus of the fusion polypeptide.

It is noted that the N-terminal apoprotein sequence usually is linked to another, linking, immunologically inert amino acid residue sequence, which extends to the amino-terminus of the complete expressed polypeptide molecule. The linking amino acid residue sequence is typically an artifact of the expression system employed to synthesize the polypeptide and is immunologically "inert", i.e., it does not substantially interfere with the ability of the fusion polypeptide to immunoreact with pan anti-apolipoprotein antibodies. An exemplary immunologically "inert" amino acid residue sequence in this regard is a  $\beta$ -galactosidase moiety as generated by a pUR expression vector.

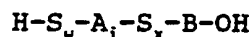
When an amino acid residue sequence other than those of a pan apo B-100 epitope or a pan apo A-I epitope is provided at the N-terminus of a subject polypeptide, the resulting molecule is itself a fusion polypeptide even though it may contain only one of the apo A-I or apo B-100 epitope amino acid residue

sequences. The N-terminal linking sequence in this case is preferably immunologically inert.

5 However, a linking sequence can in certain embodiments inhibit the interaction of the polypeptide with antibodies to the polypeptide. In the latter case, it is preferred that the interfering region of the polypeptide, which causes the inhibition, is cleavable from the targeted region of the molecule.

10 Cleavage of the interfering region can be facilitated by the introduction of a "hinge" region into the molecule. The hinge region defines a cleavage site between the N-terminal sequence desired to be cleaved and the targeted region of the polypeptide. Methods for introducing such a hinge  
15 region into a fusion polypeptide are described in U.S. Patent No. 5,013,653, the disclosure of which is incorporated herein by reference.

From a different vantage, a subject polypeptide, including a fusion polypeptide, is  
20 conveniently represented by the following formula:



wherein:

A<sub>i</sub>, when present, represents a pan apo A-I epitope amino acid residue sequence, and B represents  
25 a pan apo B-100 epitope amino acid residue sequence; and

S<sub>w</sub> and S<sub>x</sub>, when present, each represents a spacing (linking) immunologically inert amino acid residue sequence. Each of variables i, w and x, can  
30 vary independently of each other and can equal zero or one. When a variable is equal to zero, the amino acid residue sequence with which it is associated (to the left side) is absent; and when a variable is equal to one, the sequence with which it is associated is  
35 present.



Thus, at least a before-described apo B-100 polypeptide that contains a monoclonal MB47-binding epitope is present, and a before-defined amino acid residue sequence of apo A-I can also be present. When  
5 an "S" amino acid residue sequence is present in a fusion polypeptide, as represented by the above formula, the "S" sequence is as defined hereinbefore for a linking amino acid residue sequence, which can also include a portion of an immunologically inert,  
10 protein that is an artifact of expression.

Accordingly, a particularly preferred fusion polypeptide, an apo A-I/B-100 fusion polypeptide, has  $i=1$  wherein the "A<sub>i</sub>" sequence closer to the N-terminus (H) of the molecule is an amino acid residue sequence  
15 of apo A-I containing a pan epitope, and the B sequence closer to the C-terminus (OH) of the molecule is an amino acid residue sequence of a subject apo B-100 polypeptide discussed before. Also, a preferred apo A-I/B-100 fusion polypeptide has  $w=1$  and  $x=1$  in  
20 which case the "S" sequence, S<sub>w</sub>, includes an amino acid residue sequence of  $\beta$ -galactosidase, present as an artifact of expressing the fusion polypeptide from a pUR291 plasmid vector, and S<sub>x</sub> is an artifact due to restriction sites or their residual bases used in  
25 creating the expression vector, as described hereinafter.

Typically a subject polypeptide, including a fusion polypeptide, is not glycosylated; i.e., as when it is expressed by a prokaryotic host. However, a  
30 polypeptide of the invention can be eukaryotically produced, in which case it may be glycosylated. Also, a subject polypeptide can incorporate a variety of changes, such as insertions, deletions, and conservative substitutions of amino acid residues as  
35 can be present in allelic variants, as long as the

resulting molecule exhibits the desired properties. The "desired properties" as referred to herein include that the modified polypeptide has the same or similar dispersibility and immunoreactive properties as those for the unaltered polypeptide, as described  
5 hereinbefore.

When an instant polypeptide incorporates a conservative substitution in an amino acid residue sequence including the sequences presented above, the  
10 substituted amino acid residue is replaced by another, biologically similar amino acid residue such that the resulting polypeptide has an amino acid residue sequence that is compositionally different from the unsubstituted amino acid residue sequence. Some  
15 contemplated examples of conservative substitutions include substitution of a hydrophobic residue such as isoleucine, valine, leucine or methionine for another hydrophobic residue. Also, a polar residue such as arginine, lysine, glutamic acid, aspartic acid,  
20 glutamine, asparagine, and the like, can be conservatively substituted for another member of this group.

Still another aspect of a polypeptide incorporating conservative substitutions occurs when a  
25 substituted amino acid residue replaces an unsubstituted parent amino acid residue. Examples of substituted amino acids can be found at 37 C.F.R. §1.822(b)(4), which species are incorporated herein by reference. When a subject polypeptide has an amino  
30 acid residue sequence corresponding to the sequence for a native apoprotein, but for one or more conservative substitutions, preferably no more than about 40 percent, and more preferably no more than about 20 percent, of the amino acid residues of the  
35 native apoprotein are substituted in the polypeptide.

(

A subject polypeptide incorporating one or more conservative substitutions can immunologically mimic a natural allelic variant of a native apolipoprotein discussed hereinbefore. Thus, an immunologic probe is contemplated in an assay for the allelic variant.

A polypeptide of the present invention can be synthesized by any of the peptide synthetic techniques known to those skilled in the art. A summary of some of the techniques available can be found in J.M. Stuard and J. D. Young, "Solid Phase Peptide Synthesis", W. H. Freeman, Co., San Francisco (1969), J. Meinhofer, "Hormonal Proteins and Peptides" Vol. 2, pp. 46, Academic Press (New York) 1983, and U.S. Patent No. 4,631,211, which description is incorporated herein by reference. These techniques usually employ a solid phase technique such as that described in Merrifield, JACS, 85:2149 (1963). Alternatively, a subject polypeptide can be prepared by direct coupling of two smaller polypeptides, e.g., via a peptide linkage or disulfide bridge.

Preferably, however, a subject polypeptide is synthesized by recombinant DNA techniques. Such recombinant techniques are favored especially when the desired polypeptide is relatively long, e.g., more than about 50 residues in length, as is the case here for an apo B-100 polypeptide. When recombinant DNA techniques are employed to prepare a subject polypeptide, a DNA segment coding for the desired polypeptide or precursor to the polypeptide is ligated to a preselected vector for subsequent expression.

A preferred subject polypeptide is expressed by the SURE E. coli cell line (Stratagene Cloning Systems; La Jolla, CA). Thus, a conservatively substituted polypeptide incorporating any post-

translation modifications effected by SURE E. coli cells is also contemplated. Accordingly, a subject polypeptide can be expressed by SURE E. coli transfected with plasmid "84" or "144", which plasmids  
5 have a molecular weight of about 6 Kb (kilobases) as determined by agarose gel electrophoresis, can have an amino acid residue sequence conservatively substituted from that described before. A subject apo A-I/B-100  
10 fusion polypeptide can be expressed by SURE E. coli transfected with plasmid "137", which plasmid has a molecular weight of about 6.6 Kb as determined by agarose gel electrophoresis.

#### C. DNA Segments

15 A DNA segment of the invention ( a subject DNA segment) encodes a before-described subject polypeptide. A subject DNA segment is contemplated to be a discrete, "isolated" chemical species available for manipulation, as when generated by the polymerase  
20 chain reaction (PCR), as described below. All DNA segments discussed herein have both 5' and 3' ends.

Alternatively, an isolated, subject DNA segment can be operatively ligated to a vector, as when it is to be cloned or subsequently expressed in a  
25 suitable host. The DNA segment is "operatively ligated" to a vector when it is covalently bonded in-frame for expression of a subject polypeptide or fusion polypeptide to the vector, as is well known. Additionally, when the DNA segment is operatively  
30 ligated to an expression vector, a promoter sequence is present so that expression of the DNA segment is under the control of the promoter sequence. Vectors containing an operatively ligated subject DNA segment are discussed in greater detail hereinafter.

A preferred isolated DNA segment comprises a nucleotide sequence of at least 240 base pairs encoding a subject polypeptide, such as the amino acid residue sequence of apo B-100 shown in Figure 1 from about residue 3430 through about residue 3510 (e.g., the 243 bp DNA that encodes polypeptide s1 in Tables 1 or 2). The nucleotide sequence encoding the apo B-100 amino acid residue sequence can include up through about 1130 nucleotide base pairs of the apo B-100 sequence shown in SEQ ID NO:2, such as the about 1130 bp segment used to express polypeptide s9. Preferably, the apo B-100 isolated DNA segment encodes an amino acid residue sequence which is one listed in Table 1, before, and the DNA segment comprises a nucleotide sequence from the positions therein. A particularly preferred isolated DNA segment encodes apo B-100 epitope polypeptide s5, s6, s7, s8 or s9.

Also contemplated is an isolated DNA segment that encodes an amino acid residue sequence of apo A-I shown in SEQ ID NO:4 and Figure 2, including the sequence from amino acid residue 120 to residue 135: LysValGlnProTyrLeuAspAspPheGlnLysLysTrpGlnGluGlu (SEQ ID NO:5). More preferably, the entire DNA segment comprises up through about 696 nucleotides and encodes an amino acid residue sequence of apo A-I as shown in Figure 2 that encompasses the entire apo A-I molecule. Most preferably, an apo A-I-encoding DNA having about 663 bp that encodes amino acid residue positions from about residue 19 through residue 240. Any non-apo A-I residues can be selected from the full set of amino acid residues defined hereinbefore, with the previously noted caveat as to non-inhibition with the immunoreactivity of the expressed polypeptide.

An above-described DNA segment encoding an amino acid residue sequence of one apolipoprotein can

also be operatively ligated; i.e., ligated in-frame, to another DNA segment encoding a different apolipoprotein. Thus, a DNA segment comprising nucleotide sequences encoding a pan apo B-100 epitope sequence as well as a pan apo A-I epitope sequence is contemplated. A particularly preferred DNA segment containing both a pan apo B-100 epitope sequence and a pan apo A-I DNA epitope sequence is present in plasmid "137" and can be isolated as an approximately 1150-1160 bp restriction fragment having a BamHI, SalI or PstI restriction site (as are present in the polylinker of plasmid vector pUR291 discussed hereinafter) at the 5' end and a Pst I site at the 3' end. It is to be understood that longer DNA segments are contemplated as where DNA encoding polypeptides s6-s9 are utilized along with the 663 bp apo A-I DNA, as are shorter DNA segments as where a DNA segment coding for polypeptide s5 is used along with apo A-I DNA coding for the apo A-I polypeptide from residue 120 through residue 135 discussed above.

A subject isolated DNA segment or that segment ligated into a vector as a recombinant DNA molecule discussed hereinafter can also include codons that encode amino acid residues other than those of an apo B-100 or apo A-I sequence. Those other codons and encoded amino acid residues of an expressed polypeptide constitute the previously discussed spacing (linking) sequence of an  $S_{w-x}$  of the previously discussed equation.

Thus, a subject polypeptide of the previously discussed formula can be encoded by a subject isolated DNA segment. Frequently, as in the case of an expressed fusion polypeptide, the  $S_w$  portion of the previous formula is contributed by the vector, although by appropriate selection of vectors and

restriction sites, an isolated DNA segment of the invention can also encode a desired polypeptide without the spacing amino residue sequence  $S_w$ .

5 The total length of an isolated DNA segment of the invention is mostly a matter of choice once at least a before-mentioned pan apo B-100 epitope-encoding sequence is present. For example, one to several thousand base pairs can be present downstream of the expression stop codon. Similarly, one to one  
10 thousand or more base pairs can be present upstream from the apo B-100 epitope expression start codon, so long as the expression frame is maintained, as where a subject DNA segment includes codons for an apo B-100 epitope as well as a portion of the  $\beta$ -gal gene and the  
15 regulatory sequences for the  $\beta$ -gal gene.

A minimal length for such a segment is about 280 bp that encodes polypeptide s1 and the before-described 16-mer apo A-I polypeptide from amino acid residue position 120 to position 135. As a  
20 polypeptide s5-s9 is a more preferred apo B-100 polypeptide, a more preferred DNA segment at least contains a polypeptide s5-encoding sequence of about 280 bp or more preferably still the s8-encoding sequence of about 480 bp plus the 663 bp that encode  
25 the apo A-I amino acid residue sequence discussed before. The presence of additional base pairs is also contemplated for linking sequence, as discussed before.

An RNA segment equivalent (complementary) to a  
30 DNA segment described above is also contemplated. In this embodiment, an RNA segment can be ligated to an RNA vector, e.g., viral RNA, as is well known.

A subject DNA segment can be readily synthesized by chemical techniques, e.g., by the well-  
35 known phosphotriester method [Matteuci et al., JACS,

103:3185 (1981)]. By chemically synthesizing the DNA segments, any desired substitution, insertion or deletion of an amino acid residue or sequence from a template polypeptide can be readily provided by making  
5 the corresponding change in the nucleotide sequence of the DNA segment.

In addition to synthetic chemical techniques, a DNA segment of the invention can be produced by enzymatic techniques. Thus, restriction enzymes,  
10 which cleave DNA molecules at predefined recognition sequences, can be used to isolate DNA fragments from larger DNA molecules that encode a subject polypeptide. Typically, DNA fragments produced in this manner have cohesive, "overhanging" termini,  
15 whereby single-stranded nucleotide sequences extend beyond the double-stranded region of the molecule. The presence of such cohesive termini is often preferred over blunt-ended DNA molecules, although blunt-ended DNA can be used as can fragments having  
20 one blunt end and one overhanging end. The isolated DNA fragments can then be operatively ligated (cloned) into a suitable vector, thereby incorporating the subject DNA segment into the vector.

Whenever an RNA segment coding for a subject  
25 polypeptide is used, an RNA molecule including the RNA segment is transcribed into complementary DNA (cDNA) via a reverse transcriptase. The cDNA molecule can then be ligated to an expression vector which can be transfected in a suitable host to give the desired  
30 subject polypeptide.

A DNA segment coding for an above-described amino acid residue sequence can be provided with start and stop codons or one or both of the start and stop  
35 codons can be provided by the larger DNA molecule, e.g., vector, operatively ligated to the DNA segment.



A desired polypeptide of the invention can be prepared by judicious placement of the start and stop codons. For instance, a nucleotide sequence can be provided at one or both of the 3' and 5' ends of the DNA segment  
5 so that a polypeptide is expressed which is larger than a pan epitope amino acid residue sequence discussed before by inclusion of an N-terminal or C-terminal amino acid residue sequence. Additionally, it is contemplated that regulator, promoter and  
10 operator loci can be provided at the 5' end of a subject DNA segment as desired.

In addition to standard chemical or enzymatic synthetic techniques, an instant DNA segment can be generated using a polymerase chain reaction (PCR)  
15 protocol. In PCR, a specific polynucleic acid target is transcribed by a reaction in which a primer molecule complementary to a particular section of a nucleic acid template is used to form an extension product of the primer including a nucleic acid region  
20 complementary to the target. After separation of template and extended primer, each primer extension product acts as a template and specifically anneals with a complementary primer molecule. The resulting primed template acts as a substrate for further  
25 extension reactions. These steps are repeated, preferably using an automated cycling procedure, thereby exponentially amplifying the initial polynucleic acid target to which the primer hybridizes. Procedures for conducting PCR have been  
30 extensively described, see, e.g., U.S. Patent Nos. 4,683,195 and 4,683,202, which descriptions are incorporated herein by reference.

As a target DNA segment frequently is present only in very small quantities, PCR affords sufficient  
35 amplification of the target DNA segment to permit

efficient cloning and expression of the targeted segment. For instance, plasmids previously cloned with an apo B-100 cDNA segment that includes the targeted template DNA region can be used in PCR. A  
5 subject DNA segment can thereby be generated as a discrete species.

A preferred template plasmid coding for the 3214 to 3727 amino acid residue sequence of apo B-100 is designated "1.5Kb" and is available from Dr.  
10 Stephen Young of the Gladstone Foundation Laboratories, San Francisco, CA. The "1.5Kb" plasmid is also described by Pease et al., J.Biol.Chem., 265:553 (1990). A preferred template plasmid coding for the entire amino acid residue sequence of apo A-I  
15 is designated "0.6Kb" and is available from Dr. Jan Breslow, Rockefeller University, New York, NY and from the American Type Culture Collection (ATCC) under Accession No. 57025.

As PCR employs forward and reverse single  
20 stranded oligonucleotide primer molecules to initiate the polymerization reaction at each amplification stage, forward and reverse primer molecules complementary to at least a portion of the target DNA segment are contemplated. Also contemplated are  
25 primer molecules having a nucleotide sequence that defines a recognition sequence of a preselected restriction enzyme which flanks the region of the primer molecule complementary to the target DNA segment.

30 The primer must be sufficiently long to prime the synthesis of extension products in the presence of the agents for polymerization. The exact lengths of the primers depend on many factors, including temperature and the source of primer. For example,  
35 depending on the complexity of the template sequence,

a polynucleotide primer typically contains 15 to 25 or more nucleotides, although it can contain fewer nucleotides. As few as 8 nucleotides in a polynucleotide primer have been reported as effective  
5 for use [Studier et al, Proc. Natl. Acad. Sci. USA, 86:6917-21 (1989)]. Short primer molecules generally require lower temperatures to form sufficiently stable hybridization complexes with template to initiate primer extension.

10 Forward (F) and reverse (R) primers that can be used to generate a cDNA segment coding for an above-described apo B-100 amino acid residue sequence from a "1.5 Kb" template are depicted below in Table 3. Preferred pairs of forward and reverse primers for  
15 generating a particular DNA segment are indicated in Table 4, thereafter, along with the number of the resulting plasmid in which the PCR-prepared DNAs was ligated. The primers are selected so as to define a PstI, BamHI or XhoI restriction site on either side of  
20 the targeted DNA segment.

- 50 -

Table 3  
Useful Apo B-100 Primers

	Primer No.*	Sequence
5	37F	(10,059) 5' ATGC GGATCC <u>AAATTAGAGGGCACCACAAGA</u> 3' (BamHI) (SEQ ID NO:7)
10	34F	(10,158) 5' ATGC GGATCC <u>ACTGTGAGCTTAACCACGAAA</u> 3' (BamHI) (SEQ ID NO:8)
15	33F	(10,254) 5' ATGC GGATCC <u>AATGAATACCAAGTCAAAA</u> 3' (BamHI) (SEQ ID NO:9)
20	46F	(10,287) 5' ATGC GGATCC <u>TCCTCCATGGAATTTAAGTATGAT</u> 3' (BamHI) (SEQ ID NO:10)
25	132F	(10,287) 5' ATGC CTCGAG <u>TCCTCCATGGAATTTAAGTATGAT</u> 3' (XhoI) (SEQ ID NO:11)
30	99R	(10,530) 5' ATAG CTGCAG <u>TTTGGAAGTGCCCTGGAGCTTCACTG</u> (PstI) AGACCG 3' (SEQ ID NO:12)
35	130R	(10,770) 5' TTAA CTGCAG <u>CTATTA</u> <u>GCCAAGGTCAGGGAAATCATG</u> 3' (PstI) stop (SEQ ID NO:13)
40	133R	(10,695) 5' AATT CTGCAG <u>TTATCA</u> <u>GCCGGGAGAGAGTTCCAGGGT</u> 3' (PstI) stop (SEQ ID NO:14)
45	134R	(10,632) 5' AATT CTGCAG <u>TTATCA</u> <u>GTGTAAGTGGTTTTTCGTACT</u> 3' (PstI) stop (SEQ ID NO:15)
50		

- 51 -

135R

(10,569)

5' AATT CTGCAG TTATCA AAAATTTTCTTTTACTTCAAG 3'  
 (PstI) stop

(SEQ ID NO:16)

5

10

\* A primer designated with a number plus "F" is a forward primer, whereas a primer designated with a number plus "R" is a reverse primer. Sequence numbering is as in Table 1. Restriction sites are named in parentheses beneath each sequence.

Table 4

15

Apo B-100 Primers and Plasmid Numbers

Primers (Forward/Reverse)	Plasmid Number	B-100 Coding Positions	Peptide Number
37F/99R	84	10,059-10,530	s4
20 34F/99R	83	10,158-10,530	s3
33F/99R	82	10,254-10,530	s2
46F/99R	97	10,287-10,530	s1
46F/130R	127	10,287-10,770	s8
46F/133R	146	10,287-10,695	s7
25 46F/134R	145	10,287-10,632	s6
46F/135R	144	10,287-10,569	s5

30

The primers used herein are selected to be substantially complementary to the strands of each sequence to be amplified. Accordingly, the primer contains a nucleotide sequence sufficiently complementary to non-randomly hybridize with its respective template strand. Therefore, the hybridizing region of the primer sequence may not be exactly complementary with the template.

35

40

For example, a non-complementary polynucleotide can be attached to the 5' end of the primer, with the remainder of the primer sequence being substantially complementary to the strand. Such noncomplementary polynucleotide can code for a site for protein binding

or permit adjustment of the reading frame of the codons. Also, a noncomplementary base can be interspersed in the primer, provided the primer sequence has sufficient complementarity with the sequence of the template strand to allow non-random hybridization to take place under hybridizing conditions, e.g., as when an inosine base is used for nonspecific coding. Primers having as little as a three-nucleotide exact match at the 3' end of the primer can be capable of specifically initiating primer extension products [Sommer, et al., Nuc. Acid Res., 17:6749 (1989)].

Preferably, the primer is provided in single-stranded form for maximum efficiency, but it can be double-stranded. If a double-stranded primer is used, the primer is first melted to separate its strands before being used to prepare extension products. Preferably, the primer is a polydeoxyribonucleotide.

Primers can be prepared by a variety of methods including de novo chemical synthesis and derivation of nucleic acid fragments from native nucleic acid sequences such as genes, alleles, introns, and codons of a genome, plasmid, or vector. Fragments can be obtained by such methods as restriction endonuclease limit digest of larger double-stranded nucleic acids and polymeric synthesis using a polymerase and a nucleic acid template. De novo chemical synthesis of a polynucleotide is generally preferred when a native DNA sequence coding for a subject polypeptide is known and can be conducted using any suitable method, such as the well-known phosphotriester or phosphodiester methods. See, for example, Narang et al, Meth. Enzymol., 68:90, (1979); Itakura et al, Ann. Rev. Biochem., 53:323-56 (1989); Brown et al, Meth. Enzymol., 68:109, (1979); and U.S. Patent No.

4,356,270, which description is incorporated herein by reference.

Derivation of a primer molecule from nucleic acids typically involves introducing a nucleic acid into an appropriate host as with a cloning vector, replication of the vector to increase the amount of cloned nucleic acid, followed by isolation of fragments or subfragments of the cloned nucleic acids. A description of techniques for subcloning nucleic acid fragments is found in Maniatis et al, Molecular Cloning: A Laboratory Manual, Cold Spring Harbor Laboratory, pp 390-401 (1982); and U.S. Patent Nos. 4,416,988 and 4,403,036, which descriptions are incorporated herein by reference.

Template nucleic acid sequences to be hybridized in the present methods can be present in any nucleic acid-containing sample so long as the sample is in a form, with respect to purity and concentration, compatible with nucleic acid hybridization reaction. Isolation of nucleic acids to a degree suitable for hybridization is generally known and can be accomplished by a variety of means. See, for example, Maniatis et al, Molecular Cloning: A Laboratory Manual, Cold Spring Harbor Laboratory (1982); and Ausubel et al, Current Protocols in Molecules Biology, John Wiley and Sons (1987).

The hybridization reaction is carried out under predefined conditions that permit a primer to hybridize to its complementary nucleic acid template. "Hybridizing conditions," and grammatical equivalents thereof, as used herein, refer to the set of incubation time periods, temperature and pH parameters that permit the primer to anneal with the template sequence, typically to form a nucleic acid duplex. Such hybridization conditions depend on factors such

as the length of the primer to be hybridized, the degree of complementarity between the primer and the template, the guanosine and cytidine content of the polynucleotide, the stringency of hybridization  
5 desired, and the presence of salts or additional reagents in the hybridization reaction admixture that may affect the kinetics of hybridization. Methods for optimizing hybridization conditions for a given hybridization reaction admixture are well known in the  
10 art but typically the hybridizing conditions involve solutions buffered to pH values between 4 and 9 at temperatures from 18°C to 75°C, preferably about 37°C through about 65°C, more preferably about 54°C, for time periods from 0.5 seconds to 24 hours, preferably  
15 about two minutes.

Hybridization can be carried out in a homogeneous or heterogeneous format as is well known. The homogeneous hybridization reaction occurs entirely in solution, in which both the primer and template  
20 sequences to be hybridized are present in soluble forms in solution. A heterogeneous reaction involves the use of a matrix that is insoluble in the reaction medium to which either the primer or template is bound. Also contemplated are the homogeneous  
25 hybridization reactions such as are conducted in the reverse transcription of isolated mRNA or viral RNA to form cDNA, dideoxy sequencing and other procedures using primer extension reactions in which primer hybridization is a first step.

30 Where the nucleic acid containing a template sequence is in a double-stranded (ds) form, it is preferably denatured as by heating or alkali treatment prior to conducting the hybridization reaction. The denaturation of the dsDNA can be carried out prior to  
35 admixture with a primer to be hybridized or can be



carried out after admixture of the dsDNA with the primer. Where the primer itself is provided as a double-stranded molecule, it can also be denatured prior to admixture or it can be denatured concurrently with the template-containing dsDNA.

The primer extension reaction is performed using any suitable method. Preferably, a molar excess (for genomic nucleic acid, usually about  $10^6:1$  primer:template) of the primer is admixed to the buffer containing the template strand. A large molar excess is preferred to improve the efficiency of the process. For polynucleotide primers of about 20 to 25 nucleotides in length, a typical ratio is in the range of 50 ng to 1  $\mu$ g, preferably 250 ng, of primer per 100 ng to 500 ng of mammalian genomic DNA or per 10 to 50 ng of plasmid DNA.

Deoxyribonucleotide triphosphates (dNTPs) dATP, dCTP, dGTP, and dTTP are also admixed to the primer extension reaction admixture in amounts adequate to support the synthesis of primer extension products. The amounts used are readily determined by the skilled practitioner.

In a typical primer extension reaction, the hybridization admixture is heated through about 90-100°C for about one to 10 minutes, preferably from one to four minutes. After this heating period the solution is allowed to cool to room temperature to permit primer/template hybridization. To the cooled mixture is added an appropriate agent for inducing or catalyzing the primer extension reaction. The synthesis reaction can occur at from room temperature up to a temperature above which the inducing agent no longer functions efficiently. Thus, for example, if DNA polymerase is used as the inducing agent, a

suitable temperature is generally no greater than about 40°C unless the polymerase is heat-stable.

The inducing agent can be any compound or system that effects synthesis of primer extension products. Suitable enzymes for this purpose include, for example, E. coli DNA polymerase I, Klenow fragment of E. coli DNA polymerase I, T4 DNA polymerase, T7 DNA polymerase, recombinant modified T7 DNA polymerase, and other DNA polymerases, reverse transcriptases, and other enzymes that facilitate the primer extension reaction.

Heat-stable (thermophilic) DNA polymerases are particularly preferred in this regard as they are thermally stable in an automated temperature cycling PCR format. Representative heat-stable polymerases are the DNA polymerases isolated from Bacillus stearothermophilus (Bio-Rad, Richmond, CA), Thermus thermophilus (FINZYME, ATCC #27634), Thermus species (ATCC #31674), Thermus aquaticus strain TV 1151B (ATCC #25105), Sulfolobus acidocaldarius, described by Bukhrashuili et al, Biochem. Biophys. Acta, 1008: 102-7 (1989) and by Elie et al, Biochem. Biophys. Acta, 951:261-7 (1988), Thermus filiformis (ATCC #43280), the polymerase isolated from Thermus flavus (Molecular Biology Resources; Milwaukee, WI), and "Vent" polymerases (New England Biolabs, Beverly, MA). Particularly preferred are Taq DNA polymerase, available from a variety of sources including Perkin Elmer Cetus, (Norwalk, CT), Promega (Madison, WI) and Stratagene (La Jolla, CA), and AmpliTaq™ DNA polymerase, a recombinant Taq DNA polymerase available from Perkin-Elmer Cetus.

PCR is typically carried out by cycling the following steps on a reaction admixture: 1) heating to form denatured single-stranded templates and primers,

2) cooling to permit hybridization of primer to template, and 3) maintenance at predefined conditions to permit formation of primer extension products. Methods and systems for amplifying a specific nucleic acid sequence by PCR are described in U.S. Patent Nos. 4,683,195 and 4,683,202, which are incorporated herein by reference, and in PCR Technology, Erlich, ed., Stockton Press (1989); Faloona et al, Methods in Enzymol., 155:335-50 (1987); and Polymerase Chain Reaction, Erlich et al, eds., Cold Spring Harbor Laboratory Press (1989).

Detection of amplified nucleic acid product can employ any of a variety of well known techniques. In a preferred embodiment, the amplified product is separated on the basis of molecular weight by gel electrophoresis and the separated products are visualized with nucleic acid specific stains. Although numerous nucleic acid specific stains exist and would be suitable to visualize the electrophoretically separated nucleic acids, ethidium bromide is preferred.

Alternative methods for detecting the amplified nucleic acid product include hybridization-based detection means that use a labeled oligonucleotide probe capable of hybridizing to the amplified product. Exemplary of such detection means include the Southern blot analysis, ribonuclease protection analysis using in vitro labeled polyribonucleotide probes, and similar methods for detecting nucleic acids having specific nucleotide sequences. See, for example, Ausubel et al., Current Protocols in Molecular Biology, John Wiley & Sons, 1987.

In another approach for detecting the presence of a specific nucleic acid sequence, the deoxyribonucleotide triphosphates (dNTPs) used in the

primer extension reaction include a label or indicating group that renders a primer extension product detectable. Typically such labels include radioactive atoms, chemically modified nucleotide bases, and the like. Suitable radioactive labels include  $^3\text{H}$ ,  $^{14}\text{C}$ ,  $^{32}\text{P}$ , and  $^{35}\text{S}$ , and the like.

Alternatives to radioactively labeled dNTPs are dNTPs modified chemically to contain metal complexing agents, biotin-containing groups, fluorescent compounds, and the like. A useful metal complexing agent is a lanthanide chelate compound formed by a fluorescent lanthanide metal and beta-diketonate ligands. The lanthanide binds to the nucleic acid so that a fluorescent lanthanide/nucleic acid complex is formed. See U.S. Patent Nos. 4,374,120, 4,569,790 and published International Patent Application Nos. EP0139675 and WO87/02708, which descriptions are incorporated herein by reference.

Biotin or acridine ester-labeled oligonucleotides and their use in polynucleotides have been described. See U.S. Patent No. 4,707,404, published International Patent Application EP0212951 and European Patent No. 0087636. Useful fluorescent markers for oligonucleotides include fluorescein, rhodamine, Texas Red, NBD and the like.

Techniques for separating non-hybridized labeled probes from hybridized probes are well known, and typically involve the separation of single stranded from double stranded nucleic acids on the basis of their chemical properties. Frequently, a heterogeneous separation format is used in which non-hybridized probe is separated as by washing from the labeled DNA duplexes bound to an insoluble matrix. Exemplary of this technique are Southern blots in

which the matrix is a nitrocellulose sheet and the label is  $^{32}\text{P}$  [Southern, J. Mol. Biol., 98:503 (1975)].

D. Recombinant DNA Molecules

5 A recombinant DNA molecule comprising a vector operatively ligated to an above-described DNA segment is also contemplated. The recombinant DNA molecule (DNA construct) can facilitate amplification and/or expression of a subject DNA segment coding for a polypeptide of the invention.

10 A contemplated recombinant DNA molecule incorporates a cDNA segment coding for an amino acid residue sequence of a pan native apo B-100 epitope. Preferably, the apo B-100 cDNA is selected from those identified above in Table 1 with reference to the apo B-100 sequence depicted in Figure 1 and in SEQ ID NO:2.

20 A particularly preferred recombinant DNA molecule, which is replicable upon transfection in a suitable host cell, contains a subject DNA segment encoding the amino acid residue sequence of apo B-100 shown in Figure 1 from about residue 3430 through about residue 3510, corresponding to residue 217 through residue 297 of SEQ ID NO:1. A more preferred DNA construct encodes the apo B-100 amino acid residue sequence from about residue 3430 through about residue 3520, corresponding to residue 217 through residue 307 of SEQ ID NO:1. Preferably, the DNA construct contains a nucleotide sequence up through about 1130 base pairs encoding a subject polypeptide.

25 Typically, a recombinant DNA molecule of the present invention contains a DNA sequence coding for an immunologically "inert" linking peptide sequence fused to an above-described apo B-100 amino acid residue sequence. An exemplary sequence in this

35

regard codes for a  $\beta$ -galactosidase moiety. Such a recombinant DNA molecule can be obtained by inserting a subject DNA segment into vector containing a gene encoding the  $\beta$ -galactosidase protein.

5           A contemplated recombinant DNA molecule (plasmid) can be circularized or linearized. An exemplary linearized molecule in this regard contains a before-described subject DNA segment encoding apo B-100. Similarly contemplated is a recombinant DNA  
10 molecule containing a nucleotide sequence coding for an amino acid residue sequence of apo A-I, which DNA molecule is operatively ligated in frame to the amino-terminus of a subject DNA segment encoding an apo B-100 sequence. Accordingly, a preferred linear  
15 recombinant DNA molecule comprises a nucleotide sequence encoding an amino acid residue sequence of apo A-I as shown in SEQ ID NO:3 from about amino acid residue 120 through about amino acid residue 135. More preferably, a plasmid encodes the amino acid  
20 residue sequence of apo A-I from about amino acid residue 19 through about amino acid residue 240. In the latter example, the nucleotide sequence encoding the apo A-I sequence contains about 663 nucleotides, or as the utilized double stranded molecule, 663 bp.

25           A further aspect of the invention relates to a recombinant DNA molecule that includes a DNA segment coding for an A-I/B-100 fusion polypeptide. The vector molecule permits amplification and/or  
30 expression of a fusion polypeptide that includes a before-described amino acid residue sequence of apo A-1 operatively linked to a before-described, subject amino acid residue sequence of apo B-100.

Accordingly, an exemplary vector encoding a subject apo A-I/B-100 fusion polypeptide comprises:

(a) a first nucleotide sequence up through about 696 nucleotides in length that encodes an amino acid residue sequence including the amino acid residue sequence of the pan native apo A-I epitope shown in SEQ ID NO:3 from about amino acid residue 120 through about residue 135, with the second nucleotide sequence operatively ligated to the first nucleotide sequence; and

(b) a second nucleotide sequence up through about 1130 nucleotides in length that encodes an amino acid residue sequence including the amino acid residue sequence of the pan native apo B-100 epitope shown in Figure 1 from about amino acid residue 3430 through about amino acid residue 3510 corresponding to residues 217 through 297 of SEQ ID NO:1.

The choice of vector to which a DNA segment of the present invention is operatively ligated to form a subject recombinant DNA molecule depends on the functional properties desired, e.g., efficiency of transcription, efficiency of expression within the selected transformation host cell, location of restriction sites (as when PCR products are directly cloned into the vector), and the like. However, a vector of the present invention is at least capable of directing the replication of a DNA segment coding for a subject polypeptide.

Preferably, a chosen vector includes a prokaryotic replicon; i.e., a DNA sequence having the ability to direct autonomous replication and maintenance of the recombinant DNA molecule extrachromosomally in a prokaryotic host cell transformed therewith. Such replicons are well known in the art. In addition, a vector that includes a prokaryotic replicon preferably also includes a drug resistance gene so that hosts transformed with a

vector can be readily screened. Typical bacterial drug resistance genes are those that confer resistance to ampicillin or tetracycline.

Vectors that include a prokaryotic replicon preferably include a prokaryotic promoter capable of directing the transcription of the instant polypeptide genes. A promoter is an expression control element formed by a DNA sequence that promotes binding of RNA polymerase and transcription of single-stranded DNA into messenger RNA (mRNA) molecules. Promoter sequences compatible with bacterial hosts, such as a tac promoter, are typically provided in plasmid vectors having convenient restriction sites for insertion of a DNA segment of the present invention. Typical of such vector plasmids are pUC8, pUC9, pBR322 and pBR329 available from Biorad Laboratories, (Richmond, CA) and pPL, pUR, and pKK223 available from Pharmacia (Piscataway, NJ).

Expression vectors compatible with eukaryotic cells, preferably those compatible with vertebrate cells, can also be used to form a recombinant DNA molecule described before. Eukaryotic cell expression vectors are well known in the art and are available from several commercial sources. Typically, such vectors are provided with convenient restriction sites for insertion of the desired DNA segment. Typical of such vectors are pSVL and pKSV-10 (Pharmacia), pBPV-1pML2d (International Biotechnologies, Inc.), and pTDT1 (ATCC, #31255). A preferred drug resistance marker for use in vectors compatible with eukaryotic cells is the neomycin phosphotransferase (neo) gene. [Southern et al., J. Mol. Appl. Genet., 1:327-341 (1982)].

Retroviral vectors that express a recombinant DNA of the present invention are also contemplated.



The construction and use of retroviral vectors for expressing desired DNA molecules have been described. See, e.g., Sorge, et al., Mol. Cell. Biol., 4:1730-37 (1984).

5           A number of methods are available to  
operatively link an instant DNA segment to a vector  
via complementary cohesive termini. For instance,  
complementary homopolymer tracts can be added to the  
DNA segment to be inserted and to the vector DNA. The  
10       vector and DNA segment are then hybridized by hydrogen  
bonding between the complementary homopolymer tails to  
form recombinant duplex DNA molecules. Alternatively,  
synthetic linkers containing one or more restriction  
sites can be used to join an instant DNA segment to a  
15       vector.

When a DNA segment is prepared by endonuclease  
restriction digestion, it can be treated with a DNA  
polymerase that removes protruding 3' single-stranded  
termini and fills in recessed 3' ends, thereby  
20       generating blunt-ended DNA segments. Blunt-ended DNA  
segments are incubated with a large molar excess of  
linker molecules in the presence of an enzyme, such as  
bacteriophage T4 DNA ligase, which is able to catalyze  
the ligation of blunt-ended DNA molecules. Thus, the  
25       products of the reaction are DNA segments covalently  
joined at their ends to linker sequences having  
restriction sites therein. The restriction sites of  
these DNA segments are then cleaved with the  
appropriate restriction enzyme and the segments  
30       ligated to an expression vector having termini  
compatible with those of the cleaved DNA segment.  
Synthetic linkers containing a variety of restriction  
endonuclease sites are commercially available from a  
number of sources including International  
35       Biotechnologies, Inc. (New Haven, CT).

Preferably, a subject DNA segment is ligated to a vector selected from the pUR class of vectors. See, Sambrook et al., Molecular Cloning - A Laboratory Manual, 2nd ed., Cold Spring Harbor Laboratory Press, 5 (1989) pp 17.4-5. A particularly preferred vector for cloning a subject DNA segment, which encodes a truncated acid residue sequence of apo B-100, is pUR291.

A preferred and more preferred recombinant DNA 10 molecule (expression vector) for transformation and expression in SURE E. coli of a subject polypeptide, which immunologically mimics a pan native apo B-100 epitope, are designated plasmid "84" and plasmid "144", respectively. SURE E. coli separately 15 transformed with plasmid "144" or plasmid "84" were deposited with the American Type Culture Collection (ATCC), 12301 Parklawn Drive, Rockville, Maryland USA 20852. These deposits were made on September 26, 1991 and received Accession No. 75112 and No. 75110, 20 respectively.

A preferred expression vector for transformation and expression of an above-described fusion polypeptide, which immunologically mimics a pan native apo A-I epitope and a pan native apo B-100 25 epitope, is designated plasmid "137". SURE E. coli transformed with plasmid "137" were deposited with the ATCC on September 26, 1991, and received Accession No. 75111.

The before-discussed hybridoma and the above 30 three bacterial deposits were made in compliance with the Budapest Treaty requirements that the duration of the deposits should be for 30 years from the date of deposit or for five years after the last request for the deposit at the depository or for the enforceable 35 life of a U.S. patent that matures from this

application, whichever is longer. A deposit will be replenished should it become non-viable at the depository.

5                   E.   Transformation and Purification

Also contemplated within the invention are cells stably transformed with a recombinant DNA molecule, including an expression vector, that contains an above-described DNA segment. The host  
10 cell can be either prokaryotic or eukaryotic. Preferred prokaryotic host cells are strains of E. coli, e.g., the E. coli strain SURE commercially available from Stratagene Cloning Systems (La Jolla, CA). Preferred eukaryotic host cells include yeast  
15 and mammalian cells, preferably vertebrate cells such as those from mouse, rat, goat, or primate cell lines.

The above-noted SURE strain of E. coli is particularly preferred as that strain remained stably transformed. E. coli strains DH5 $\alpha$  (BRL), JM109  
20 (Stratagene) and MC1601 (ATCC) could not be stably transformed. It is believed that the E. coli transformants prepared in Pease et al., J. Biol. Chem., 265:553-568 (1990) were not stable transformants.

25                   Transformation of a suitable host cell such as SURE strain E. coli with a recombinant DNA molecule of the present invention is accomplished by well-known methods that typically depend on the type of vector used. With regard to transformation of prokaryotic  
30 host cells, see, e.g., Maniatis et al., Molecular Cloning. A Laboratory Manual, Cold Spring Harbor Laboratory, Cold Spring Harbor, NY (1982). With regard to transformation of vertebrate cells with retroviral RNA vectors, see, e.g., Sorge et al., Mol. Cell. Biol., 4:1730-37 (1984).  
35

Successfully transformed cells; i.e., those stably transfected with a recombinant DNA molecule of the present invention, can be identified by well-known techniques. For example, stably transformed cells can be cloned to produce monoclonal colonies. Cells from those colonies can be harvested, lysed and their DNA content examined for the presence of the desired DNA segment using a method such as that described by Southern, J. Mol. Biol., 98:503 (1975).

In addition to directly assaying for the presence of the desired DNA segment, successful transformation can be confirmed by well-known immunological methods when the transformed cells express an above-described polypeptide. In this method, samples of cells suspected of being transformed are maintained under culture in an appropriate nutrient culture medium for a time period sufficient to express the polypeptide, harvested and assayed for a desired antigenicity using antibodies that immunoreact with (specifically bind to) a subject polypeptide. Preferred antibodies in this regard are produced by the hybridomas MB47 (ATCC HB 8746) when assaying for apo B-100 antigenicity and AI-11 (ATCC HB 9201) when assaying for apo A-I antigenicity.

Also contemplated are cultures of stably transformed host cells. Nutrient media useful for culturing transformed host cells are well known in the art and can be obtained from several commercial sources. When a mammalian host cell is employed a "serum-free" medium is preferably used.

Once stably transformed host cells are identified, those cells can be used in a method for expressing a subject polypeptide by maintaining (culturing) those transformed cells under appropriate culture conditions for a time period sufficient for

the cells to express the polypeptide. The expressed polypeptide is preferably isolated (recovered) from the cultured cells.

5       Methods for recovering an expressed protein from a culture are well-known in the art. For instance, gel filtration, gel chromatography, ultrafiltration, electrophoresis, ion exchange, affinity chromatography and related techniques can be used to isolate an expressed protein from the culture.  
10       In addition, immunochemical methods, such as immunoaffinity, immunoadsorption, and the like, can be performed using well-known methods, as exemplified by the methods described herein.

15               F.   Diagnostic Assays

          A method for assaying the native apo B-100 level of a body fluid sample containing that protein is also contemplated within the invention. The targeted apoprotein (antigen) is detected in a body  
20       fluid sample such as a vascular fluid sample that contains apo B-100. Exemplary body fluid samples include plasma, serum or whole blood, but urine, semen, and cerebrospinal fluid can also be used. As  
25       native apo B-100 levels can be correlated to LDL cholesterol levels, the present invention affords a method for assaying the LDL cholesterol level in a body fluid sample.

          Also contemplated is a method for assaying the amount of a second apolipoprotein, native apo A-I, in  
30       a body fluid sample. Since the amount of apo A-I in the sample can be correlated to the amount of HDL cholesterol in the sample, the invention affords a method for assaying the HDL cholesterol level of the sample.

Various well known heterogeneous and homogeneous protocols can be employed in competitive and direct binding assays for the presence, and preferably amount, of a targeted native apoprotein. Preferably, a solid-phase assay format is employed, which method permits physical separation of reaction product from potentially interfering reagents. Also preferred is a competitive binding protocol, which utilizes the competition of a subject polypeptide and native apoprotein in the assayed sample for binding to an antibody immunoreactive with both the subject polypeptide and native apoprotein.

Generally, an assay method of the present invention comprises admixing a body sample, e g., human serum or plasma from a patient, in a liquid, aqueous medium such as water, normal saline, or a buffer, or plasma or other body fluid diluted therewith, with a subject polypeptide and an antibody composition containing pan anti-apo B-100 antibodies that immunoreact with the polypeptide and any native apo B-100 in the sample. The immunoreaction admixture thus formed is maintained under biological assay conditions for a time period sufficient for the antibody composition to immunoreact with antigens in the admixture; i.e., with both subject polypeptide and with native apo B-100. The presence, and preferably amount of any immunocomplex formed, as between native apo B-100 and pan anti-apo B-100 antibodies or, preferably, between a subject polypeptide and the pan anti-apo B-100 antibodies, is then determined. The amount of immunocomplex formed can then be related to the amount of native apo B-100 in the sample using well known techniques.

Along the lines described above for assaying native apo B-100 levels, a presently described method

can also be utilized in an assay for a second apolipoprotein. Such a method comprises admixing a fluid body sample with a subject fusion apo A-I/B-100 polypeptide and an antibody composition containing pan anti-apo A-I antibodies. The pan anti-apo A-I antibodies immunoreact with native apo A-I in the sample and with the apo A-I/B-100 fusion polypeptide.

The immunoreaction admixture formed above is maintained under biological assay conditions for a time period sufficient for the anti-apo A-I antibodies to immunoreact with their antigens in the admixture. The amount of any immunocomplex formed, as between the fusion polypeptide and anti-apo A-I antibodies, is then determined. The amount of immunocomplex formed can then be related to the amount of native apo A-I in the sample, as discussed above.

The maintenance time for immunoreaction to take place can vary widely, as is well known. Maintenance times typically range from about one minute to about two hours, with times of about 30 to about 90 minutes being typically utilized.

The phrases "anti-apo B-100" and "anti-A-I antibodies" and "pan anti-apo B-100 antibodies" and "pan apo A-I antibodies" are utilized herein to mean antibody preparations that immunoreact with a before-described amino acid residue sequence of a pan native apo B-100 polypeptide epitope or of a pan native apo A-I epitope, and particularly a sequence containing the before-discussed amino acid residue sequence of the apo A-I molecule from amino acid residue position 120 to residue position 135. Particularly preferred antibody compositions are the before-discussed, deposited monoclonal antibodies such as MB47 and AI-11.

Polyclonal antibodies that immunoreact with a subject apo B-100 polypeptide or with a pan native apo A-I epitope are also contemplated. Such polyclonal antibodies can be prepared by immunization with LDL particles or apo B-100, or with HDL particles or apo A-I, respectively. A previously discussed dispersible apo B-100 polypeptide such as s5-s9 can also be used as an immunogen, as can a polypeptide containing the 120-135 amino acid residue sequence of apo A-I.

An antibody preparation can contain intact antibodies, or the binding site-containing (paratope-containing) portions thereof. Exemplary paratope-containing portions include Fab, F(ab')<sub>2</sub> and F<sub>v</sub> portions as are well known. In addition, where an antibody Fc portion is present, that portion can have the amino acid residue sequence characteristic of an animal different from the paratope-containing portion. For example, a human Fc portion can be present along with a mouse binding site-containing portion as can be accomplished by genetic engineering techniques as are well known.

The "biological assay conditions" employed in the present assay methods are selected to maintain the biological activities of the sample being assayed, as well as the antibodies and polypeptides used in the assay. Typically acceptable "biological assay conditions" include a temperature in the range of about 4°C through about 45°C, preferably about 37°C; a pH value in the range of about 5 through about 9, preferably about 7; and an ionic strength ranging from that for distilled water to that of about 1M sodium chloride, preferably about that of physiological saline. Methods for optimizing such conditions are known to the skilled practitioner.



A predetermined amount of each of the various reagents is utilized in these assays. The predetermined amount of any reagent can vary with the assay. However, as the amount of apo B-100 and/or apo A-I is to be determined, the amounts of the standards such as a dispersible subject polypeptide or apo A-I/apo B-100 polypeptide must be known so that results can be quantified.

The manner of determining the presence or amount of an immunocomplex formed in a subject assay depends upon the method desired to identify the immunocomplex. For instance, an antibody used in the assay can be labeled prior to or subsequent to forming an immunocomplex with a dispersible antigenic polypeptide of the present invention or apolipoprotein. The labeled immunocomplex can be quantitated by methods appropriate for detecting the respective label, e.g., radiation detection when a radioactive label is used, fluorescence detection when a fluorescent is employed, reaction with avidin when a biotin label is employed, conversion of an enzyme substrate into a detectible product when an enzyme label is used, and the like. Alternatively, an unlabeled immunocomplex can be detected by forming an immunoreaction product between the immunocomplex and a labeled substance that specifically binds the immunocomplex, e.g., a labeled anti-antibody. A label is thereby attached to the assayed immunocomplex, which affords ready detection.

A particularly preferred method for assaying for the amount of native apo B-100 in a body fluid sample containing (or suspected of containing) that protein employs a dispersible subject polypeptide (antigen) affixed to the surface of a solid support to provide a "solid-phase" polypeptide. (Methods of

affixation are discussed hereinafter.) Such a method, often referred to as a competitive assay, comprises the steps (a)-(c) below:

5 (a) admixing a predetermined amount of a body fluid sample containing native apo B-100 in a liquid medium with predetermined amounts of a dispersible subject polypeptide affixed to a solid support and pan anti-apo B-100 antibodies to form a solid-liquid phase admixture. The pan anti-apo B-100 antibodies  
10 immunoreact with the native apo B-100 of the sample and with the subject polypeptide. The polypeptide includes an amino acid residue sequence about 80 through about 375 residues of apo B-100 shown in Figure 1 and includes the sequence from about residue  
15 3430 through about residue 3510 corresponding to residues 217 through 297 of SEQ ID NO:1, and preferably the sequence from about residue 3430 through about 3520 corresponding to residues 217 through 307 of SEQ ID NO:1;

20 (b) maintaining the solid-liquid phase admixture under biological assay conditions for a time period sufficient for the pan anti-apo B-100 antibodies to immunoreact with the polypeptide and apo B-100 to form a solid-phase polypeptide/anti-apo B-100  
25 immunocomplex and a liquid phase apo B-100/anti-apo B-100 immunocomplex, respectively;

(c) separating the solid and liquid phases; and  
(d) determining the amount of solid-phase polypeptide/anti-apo B-100 immunocomplex formed, and  
30 thereby the amount of native apo B-100 in the body fluid sample.

The above assay can be conducted by admixing the reagents in any order. A preferred order of admixture involves admixing the solid-phase

polypeptide with the body fluid sample prior to admixing with the pan anti-apo B-100 antibodies.

Usually, a blocking agent, which blocks non-specific binding of pan anti-apo B-100 antibodies with the solid phase matrix, is admixed with and any excess removed from the solid phase-affixed polypeptide prior to contacting the polypeptide with the anti-apo B-100 antibodies. A preferred blocking agent in this regard is BSA (bovine serum albumin), but any immunologically inert proteinaceous material can be used.

An especially preferred assay employs an enzyme-labeled MB47 antibody. A preferred method for determining the amount of solid-phase immunocomplex formed involves monitoring the reaction of enzyme-labeled MB47 antibody with a substrate for the enzyme, since the rate of such reaction depends on the amount of label affixed to the immunocomplex.

Another method for assaying the native apo B-100 level in a body fluid sample utilizes a subject polypeptide to inhibit the immunoreaction of native apo B-100 in the sample with pan anti-apo B-100 antibodies. The degree of inhibition observed can be correlated with the amount of native apo B-100 antigen in the sample by analogy to well known techniques. This method is often referred to as a competitive inhibition assay. The method comprises the steps (a)-(d) below:

(a) admixing a predetermined amount of a body fluid sample containing native apo B-100 in a liquid medium with predetermined amounts of a before-described subject polypeptide and solid phase-affixed pan anti-apo B-100 antibodies to form an admixture. The anti-apo B-100 antibodies immunoreact with the native apo B-100 of the sample and with the polypeptide;

(b) maintaining the admixture under biological assay conditions for a time period sufficient for the anti-apo B-100 antibodies to immunoreact with the polypeptide and apo B-100 to form a solid phase-  
5 affixed polypeptide/anti-apo B-100 immunocomplex and apo B-100/anti-apo B-100 immunocomplex;

(c) separating the solid and liquid phases; and  
(d) determining the amount of solid phase-  
10 affixed polypeptide/anti-apo B-100 immunocomplex formed, and thereby the amount of native apo B-100 in the body fluid sample.

A subject polypeptide is here conjugated to a label or indicating means such as one selected from the group of a radioactive element, enzyme, biotin,  
15 fluorescent, and chemiluminescent labels. The amount of indicating mean or label bound can then be used to determine the amount of apo B-100 in the sample.

A subject polypeptide can also be employed in standardizing an assay for the amount of native apo  
20 B-100 in a body fluid sample. In this procedure, a known amount of a subject polypeptide can be quantitated with an indicating means, such as a labeled antibody, and a body fluid sample containing an unknown amount of apo B-100 can be quantitated with  
25 the same indicating means. When these assay results are correlated, the unknown amount of native apo B-100 is determined. This standardization process can be used with any apo B-100 assay described herein.

The standardization method comprises the  
30 following steps:

(a) admixing in a liquid medium first predetermined amounts of a body fluid sample containing native apo B-100 and pan anti-apo B-100 antibodies to form a first admixture, in which the

anti-apo B-100 antibodies immunoreact with the native apo B-100 of the sample;

(b) maintaining the first admixture under biological assay conditions for a time period  
5 sufficient for the anti-apo B-100 antibodies to immunoreact with the native apo B-100 in the sample to form a native apo B-100/anti-apo B-100 immunocomplex;

(c) admixing second predetermined amounts of the same pan anti-apo B-100 antibodies and a subject  
10 polypeptide in a liquid medium to form a second admixture in which the anti-apo B-100 antibodies immunoreact with the polypeptide;

(d) maintaining the second admixture under biological assay conditions for a time period  
15 sufficient for the anti-apo B-100 antibodies to immunoreact with the polypeptide to form a polypeptide/anti-apo B-100 immunocomplex; and

(e) determining the amounts of native apo B-100/anti-apo B-100 and polypeptide/anti-apo B-100  
20 immunocomplexes formed, and thereby the amount of native apo B-100 in the body fluid sample.

Preferably, the anti-apo B-100 antibodies of the above method are conjugated to a label selected from the group consisting of radioactive, enzyme,  
25 biotin, fluorescent, and chemiluminescent labels. It is also preferred that the anti-apo B-100 antibodies, be affixed to a solid support.

In another aspect of a standardization assay, the determination step (e) comprises:

(i) admixing the native apo B-100/anti-apo B-100 immunocomplex in a liquid medium with second  
30 antibodies immunoreactive with the native apo B-100 to form a third admixture, in which the second antibodies are operatively linked to a first label;

(ii) maintaining the third admixture under biological assay conditions for a period of time sufficient for the second antibodies to immunoreact with the native apo B-100/anti-apo B-100 immunocomplex to form a second antibody/native apo B-100/anti-apo B-sandwich complex;

(iii) determining the amount of first label affixed to the second antibody/native apo B-100/anti-apo B-100 sandwich complex, thereby determining the relative amount of native apo B-100/anti-apo B-100 immunocomplex formed;

(iv) admixing the polypeptide/anti-apo B-100 immunocomplex in a liquid medium with third antibodies immunoreactive with the polypeptide or the bound antibodies to form a fourth admixture, the third antibodies operatively linked to a second label;

(v) maintaining the fourth admixture under biological assay conditions for a period of time sufficient for the third antibodies to immunoreact with the polypeptide/anti-apo B-100 immunocomplex to form a third antibody/polypeptide/anti-apo B-100 sandwich complex; and

(vi) determining the amount of second label affixed to the third antibody/polypeptide/anti-apo B-100 sandwich complex, thereby determining the amount of polypeptide/anti-apo B-100 immunocomplex formed and the true amount of native apo B-100 in the sample.

Preferably, the second antibody above is an MB24 antibody secreted by the hybridoma having ATCC Accession No. HB 8742 and the third antibody is an anti-beta-galactosidase antibody where a beta-galactosidase fusion protein is used as the subject polypeptide. Antibodies that immunoreact with another epitope of a subject polypeptide, as can be formed

using that polypeptide as an immunogen, and whose immunoreaction is not inhibited by the bound anti-apo B-100 antibodies can also be used as the third antibodies. Label-linked anti-MB47 antibodies such as enzyme- or radio-labeled goat anti-mouse antibodies can also be used. It is also preferred that the first and second markers are identical.

A before-described dispersible apo B-100 subject polypeptide including a fusion polypeptide is thus used as the long-awaited standard for the otherwise unstable intact, whole apo B-100 molecule.

In addition to a method for assaying native apo B-100 in a body fluid sample, a method for assaying the levels of native apo A-I in the sample is contemplated. In particular, a preferred method of assaying for the amounts of native apo B-100 and apo A-I antigens in a body fluid sample employs a subject apo A-I/B-100 fusion polypeptide (a recombinant apo A-I/B-100 antigen) affixed to the surface of a solid support. The method comprises the following steps:

(a) admixing a predetermined amount of the body fluid sample to be assayed in a liquid medium with predetermined amounts of a dispersible subject fusion polypeptide affixed to a solid support, pan anti-apo B-100 antibodies, and pan anti-apo A-I antibodies enumerated previously to form a solid-liquid admixture. The anti-apo B-100 antibodies immunoreact with the native apo B-100 of the sample and with the polypeptide, and the anti-apo A-I antibodies immunoreact with the native apo A-I of the sample and with the fusion polypeptide;

(b) maintaining the solid-liquid admixture under biological assay conditions for a time period sufficient for the anti-apo B-100 antibodies and anti-apo A-I antibodies to immunoreact with the respective

antigens of the fusion polypeptide to form solid-phase fusion polypeptide/anti-apo B-100 and fusion polypeptide/ anti-apo A-I immunocomplexes; and

- 5 (c) determining the amounts of solid-phase fusion polypeptide/anti-apo B-100 and fusion polypeptide/anti-apo A-I immunocomplexes formed, and thereby the amounts of native apo B-100 and native apo A-I in the sample.

- 10 Again, the order of admixture of the above reagents is not critical; however, the reagents are preferably admixed substantially simultaneously. The above assay can be carried out using a single solid support for both assays where the antibodies to each antigen are differently labeled. Exemplary different  
15 labels or markers are radioisotopes such as  $^3\text{H}$  and  $^{131}\text{I}$  that emit beta, and beta and gamma radiation, respectively, of different energies. These labels are used with a gated radiation counter. Another  
20 exemplary system uses horseradish peroxidase conjugated to one antibody and alkaline phosphatase linked to the other, along with their usual substrates. Here, optical densities at two different wavelengths are determined to obtain the desired data.

- 25 Another assay of the invention employs pan anti-apo B-100 antibodies immobilized on a first support and pan anti-apo A-I antibodies immobilized on a second support. When two apoproteins are assayed, the antibodies can be provided on two different supports. Alternatively, the antibodies can be  
30 affixed to the same support, in which case different detection means must be associated with each kind of antibody, or the different antibodies can be localized to different regions, e.g., cells or rows, in a grid or otherwise partitioned pattern on the surface of a  
35 solid matrix.



Also contemplated is a "contrived control" method for assaying the amount of native apo B-100 in a lipoprotein-containing body fluid sample. This method is termed "contrived control" because it employs a subject polypeptide as a reference or control in delipidated plasma in an assay for native apo B-100 so that both the assayed sample and control contain similar plasma proteins and salts and have similar viscosities.

In this assay, the delipidated body fluid sample comprises a portion of the liquid medium in which the subject polypeptide and anti-apo B-100 antibodies are admixed. Preferably, the amount of delipidated body fluid is the same as the amount of body fluid sample that is assayed. A contrived control assay can be used with any assay discussed herein that utilizes a separate subject polypeptide-containing aqueous composition, such as standardized, assay.

#### G. Diagnostic Systems

A diagnostic system, typically in kit form, is also contemplated that can be employed in carrying out the above-described assay methods. A diagnostic system of the invention comprises a container (package) that includes at least a subject polypeptide. The polypeptide is provided in an amount sufficient to perform at least one assay and is usually provided as a separately packaged reagent.

The polypeptide can be labeled, as when conjugated to a marker; however, the polypeptide can also be unlabeled. The polypeptide can be provided neat, dissolved or suspended in a suitable liquid vehicle such as a buffer, lyophilized, bound to a

solid support, or in another convenient physical form, as is readily apparent.

Another preferred kit comprises an above-described apo A-I/B-100 fusion polypeptide, pan anti-  
5 apo B-100 antibodies, and pan anti-apo A-I antibodies as discussed previously. Exemplary pan anti-apo B-100 antibodies include MB47 antibodies and exemplary pan anti-apo A-I antibodies include AI-11 antibodies.

Preferably, an indicating (marking or labeling)  
10 means is also provided that permits indication of an immunocomplex formed upon reaction of a subject polypeptide with an antibody immunoreactive with the polypeptide. In some formats, as is apparent from the before-described methods, a labelling means is  
15 preferably conjugated to a subject polypeptide. In other formats, a labelling means is preferably associated, as by conjugation, with an antibody immunoreactive with the polypeptide. Thus, a preferred kit contains a labeled antibody, such as a  
20 labeled murine antibody, that immunoreacts with another epitope of a subject polypeptide also provided in the kit. Optionally, a labeled antibody is provided that immunoreacts with the immunocomplex formed upon reaction of a subject polypeptide with an  
25 antibody immunoreactive with the polypeptide. An example of a latter-described labeling means is a labeled goat anti-mouse antibody that is useful for immunoreacting with before-described murine monoclonal antibodies such as MB47 and AI-11.

30 When a label is associated with a reagent provided in a kit of the invention, it can be selected from any of those commonly available. Exemplary labels include  $^{111}\text{In}$ ,  $^{99}\text{Tc}$ ,  $^{67}\text{Ga}$ ,  $^{125}\text{I}$ ,  $^{131}\text{I}$ , non-radioactive labels, such as biotin and enzyme-  
35 linked antibodies, chromophoric labels such as

fluorescein, phycoerythrin, rhodamine, chemiluminescent labels, and the like. Preferred enzyme labels include horseradish peroxidase (HRPO), alkaline phosphatase, and leuciferin. Preferred chromophoric labels include fluorescein and the Immune-lite chemiluminescent visualization system distributed by Bio-Rad (Richmond, CA).

Methods for linking a label to an immunoglobulin or protein are well-known. For instance, antibody molecules produced by a hybridoma can be labeled by metabolic incorporation of radioisotope-containing amino acids provided as a component in the culture medium [Galfre et al., Meth. Enzymol., 73:3-46 (1981)]. The techniques of protein conjugation or coupling through activated functional groups are particularly applicable. See, for example, Aurameas, et al., Scand. J. Immunol., Vol. 8, Suppl. 7:7-23 (1978), Rodwell et al., Biotech., 3:889-894 (1984), and U.S. Patent Nos. 4,493,795 and 3,996,345, which disclosures are incorporated herein by reference.

A diagnostic system of the invention can also include a specific binding agent. A "specific binding agent" is a chemical species capable of selectively binding a reagent species of a diagnostic system described herein, but is not itself an antibody molecule. Exemplary specific binding agents are complement proteins or fragments thereof, protein A, and the like that react with an antibody molecule when the antibody is part of an immunocomplex described above.

In preferred embodiments, a specific binding agent is labeled; however, it can be unlabeled. When the specific binding agent is not labeled, the specific binding agent is typically used as an

amplifying means or reagent. In these embodiments, a specific-binding indicating means, such as a labeled antibody, is also provided that specifically binds to the amplifying means when the amplifying means is bound to an immunocomplex described herein and provides a signal for the presence of the immunocomplex.

A diagnostic kit of the invention can be used conveniently in an "ELISA" format to detect the presence or quantity of a native apolipoprotein in a body fluid sample, such as serum, plasma or urine, using a before-described method. An "ELISA" format is an enzyme-linked immunosorbent assay that employs an antibody or antigen bound to a solid phase and an enzyme-antigen or enzyme-antibody conjugate to detect and quantify the amount of antibody or antigen present in a sample. A description of the ELISA technique is found in Chapter 22 of the 4th Edition of Basic and Clinical Immunology by D.P. Sites et al., published by Lange Medical Publications of Los Altos, CA in 1982. Various ELISA formats are also described in U.S. Patents Nos. 3,654,090; 3,850,752; 3,817,837; 3,875,011; and 4,016,043, which disclosures are incorporated herein by reference.

More preferably, an antibody or antigen reagent, including a recombinant antigen (fusion polypeptide), is affixed to a solid matrix to form a solid support, which is separately packaged in a subject diagnostic system. Also, a plurality of solid supports such as the wells of a 96-well plastic microtiter plate can be employed, as when a plurality of antibodies are employed in an above-described assay method or when a plurality of assays are to be carried out.

An antibody or antigen reagent of a diagnostic system or as used in a before-described assay method is typically affixed to a solid matrix by adsorption from an aqueous medium, although other modes of affixation well known to those skilled in the art can be used, such as covalent binding, binding by an antibody that itself is bound to the matrix, or binding of an antibody to a solid support via adsorbed protein A. Solid matrix materials useful in this regard include the derivatized cross-linked dextran available under the trademark SEPHADEX from Pharmacia Fine Chemicals (Piscataway, NJ), agarose in its derivatized and/or cross-linked form, polystyrene beads about 1 micron through about 5 millimeters in diameter available from Abbott Laboratories of North Chicago, IL, polyvinyl chloride, polystyrene, cross-linked polyacrylamide, nitrocellulose- or nylon-based webs such as sheets, strips or paddles, tubes, plates, the wells of a microtiter plate such as those made from polystyrene or polyvinylchloride, and the like. Microtiter wells in the form of strips and plates are particularly preferred.

A reagent provided in a diagnostic system described herein can be provided in solution, as a liquid dispersion or as a substantially dry powder, e.g., in lyophilized form. Where an indicating means is an enzyme, the enzyme's substrate can also be provided in a separate package of the diagnostic system. Usually, the reagents are packaged under an inert atmosphere. A solid support, such as a microtiter plate, and one or more buffers can also be included as separately packaged elements in this diagnostic assay system.

A more preferred diagnostic system, in one package, is useful for detection of native apo B-100

and native apo A-I. An exemplary kit in this regard includes the following in separate containers:

(a) a dispersible subject apo A-I/B-100 fusion polypeptide affixed to a microtiter plate;

5 (b) HRP0-labeled MB47 antibodies in lyophilized form; and

(c) non-HRP0-labeled, e.g., alkaline phosphatase labeled, AI-11 antibodies in lyophilized form.

10 Another exemplary kit includes the following in separate containers:

(a) a dispersible subject apo A-I/B-100 fusion polypeptide in lyophilized form;

(b) HRP0-labeled MB47 antibodies; and

15 (c) non-HRP0-labeled AI-11 antibodies.

Preferably, a kit, such as described above, contains a blocking agent, such as bovine serum albumin (BSA), for blocking nonspecific binding of antibodies to polypeptide. Also, a kit containing an  
20 HRP0-labeled reagent preferably includes: (i) a supply of hydrogen peroxide of known concentration; (ii) a visualizing oxidative dye precursor such as OPD (o-phenylenediamine); and (iii) a solution of a quenching agent such as 4N sulfuric acid.

25 A kit also is preferably provided with instructions for use of the packaged reagent. The instructions typically include a tangible expression describing the reagent concentration or at least one assay parameter such as the relative amounts of  
30 reagent and sample to be admixed, maintenance time periods for reagent/sample admixtures, temperature, buffer conditions, and the like.

In a further aspect of the invention, a diagnostic system contains means for recording and/or  
35 correlating data obtained in the above-described

assays. Thus, in one embodiment, data are recorded with a tangible medium, e.g., computer storage, chart paper, and the like. The data can be automatically input and stored using analog/digital (A/D) instrumentation, which is commercially available. Also, the data can be recalled and reported or displayed as desired for best presenting the instant correlations of data. Accordingly, materials for correlating data, such as instrumentation and software suitable for use with the present methods, can be provided in a presently-described diagnostic system and are contemplated as within the scope of the present invention.

A diagnostic system of the invention can be contained in a conventional package, including glass and plastic (e.g., polyethylene, polypropylene and polycarbonate) bottles, vials, plastic and plastic-foil laminated envelopes, paper, and the like.

#### 20 Examples

The following examples do not limit the scope of the invention but are presented merely to illustrate certain aspects of the present invention.

#### 25 Example 1: Mapping of MB47 epitope of apo B-100

The MB47 epitope of apo B-100 was mapped using  $\beta$ -gal fusion proteins containing the polypeptide of interest. This was effected by cloning PCR-amplified apo B-100 cDNA into a polycloning site located at the 3' terminus of the lacZ gene of the plasmid pUR291 [Ruther et al., EMBO J., 2:1791 (1983)]. The pUR291 vector has a unique PstI site within the polycloning site. See, Sambrook et al., Molecular Cloning - A Laboratory Manual, 2nd ed., Cold Spring Harbor

Laboratory Press, pp. 17.4-5, (1989) for a restriction map of the relevant cloning site.

Exemplary polypeptide regions of apo B-100 that were mapped by this method are identified in Table 1 hereinbefore; the primers used to generate the amplified cDNAs are identified in Tables 3 and 4.

Notably, one primer set found ineffective for preparing an immunoreactive polypeptide were the oligonucleotides denoted #37 (forward) and #35 (reverse) which had the following nucleotide sequences:

5'-ATGCGGATCCAAATTAGAGGGCACCACAAGA-3'  
(#37, Forward) which is identified as SEQ ID NO:17.

5'-GTGCCCTGCAGTTATCACTTCACTGAAGACCGTGTGCT-3'  
(#35, Reverse) which is identified as SEQ ID NO:18.

These primers coded for sequence positions 3349 through 3506, nucleotide positions 10,059 through 10,770. This primer set encoded a polypeptide (3349-3506) substantially identical to positions 3351-3506 reported in [Pease et al., J. Biol. Chem., 265:553 (1990)] as apparently useful in Western blots, but exhibited no binding in assays conducted herein.

#### A. Cloning of 483 bp Apo B-100 cDNA

The procedure for cloning the PCR-amplified apo B-100 cDNA into a pUR291 vector is described hereinbelow for the 483 bp apo B-100 coding fragment identified in Table 2; however, the same general procedure was used for other domains.

Briefly, vector pUR278 (a vector similar to pUR291) containing a 1.4kb apo B-100 cDNA was obtained from R. Pease of the Medical Research Council Clinical Research Centre, Harrow HA1 3UJ, United Kingdom [Pease



et al., supra]. The apo B-100 cDNA of this vector starts at nucleotide 9639 and terminates with nucleotide 11,709 [Ludwig et al. DNA, 6:363 (1987); Knott et al., Nucl. Acids Res., 14:7501 (1986)]. This vector was used as a template for generating the apo B-100 inserts.

The pUR291 plasmid was selected to express the targeted apo B-100 fragments as  $\beta$ -gal fusion proteins. The pUR291 plasmid carries an entire active  $\beta$ -gal gene including promoter, and generally works well for preparing fusion proteins containing an N-terminal  $\beta$ -galactosidase. The 3' terminus of the  $\beta$ -gal gene has convenient BamHI, PstI and HindIII restriction sites. Also, pUR291 confers ampicillin resistance to the transformed host.

The 483 base pair (bp) apo B-100 nucleotide sequence extending from base 10,287 to base 10,770 was amplified using the 1.4kb insert of pUR278 as a template. Thus, forward and reverse primers #46 and #130, respectively of Table 3, were hybridized with pUR278 and the resulting hybrid was subjected to standard PCR conditions (Perkin Elmer Cetus; Norwich, CT). After 40 PCR cycles, the amplification reaction was halted and the product containing the amplified 483 bp nucleotide sequence was separated from the reaction mixture.

As the #46 and #130 primers (Table 3) carried BamHI and PstI restriction sites, the amplified product could be cut with those enzymes and cloned directly into the pUR291 vector, previously cut with BamHI and PstI. Although the 483 bp-containing sequence actually contained more than that number of base pairs because of the bases added to create the BamHI and PstI restriction sites, the produced fragment and the fragment resulting from cleavage with

those enzymes is referred to for convenience as a 483 bp fragment or 483 bp PCT product.

B. Cloning of Apo B-100 Sequences

5       The isolated 483 bp-containing PCR product was digested with BamHI and PstI restriction enzymes (10  $\mu$ l each) for one hour and 37°C to generate cohesive termini. The 483 bp sticky ended fragment was isolated and ligated into the BamHI- and PstI-  
10       linearized plasmid pUR291 using T4 DNA ligase (Bethesda Research Laboratory; Grand Island, NY).

15       The other recombinant apo B-100 polypeptides noted in Table 1 and Table 2, and the primers used for their preparation as noted in Tables 3 and 4, were similarly prepared, and utilized as discussed below.

C. Transformation in SURE E. coli.

20       The resulting plasmid vector was isolated and used to transform SURE E. coli following the supplier's protocol (Stratagene Cloning Systems; La Jolla, CA). Briefly, 100  $\mu$ l of the cells were aliquotted into a pre-chilled 15 ml Falcon 2059 polypropylene tube and 1.7  $\mu$ l of fresh 1:10  
25       2-mercaptoethanol were added to the cells to give a 25 mM final concentration. The reaction mixture in the tube was swirled gently and chilled in ice for 10 minutes. Approximately 0.1-50 ng plasmid were added to the cells and the mixture swirled gently. A  
30       control insert of 1  $\mu$ l of pBR322 was admixed with another 100  $\mu$ l of cells and swirled gently.

35       The tubes were placed in ice for 30 minutes then heat pulsed in a 42°C water bath for 45 seconds. The tubes were returned to ice for two minutes. About 0.9 ml of preheated (42°C) SOC medium [20 g Bacto-tryptone, 5 g yeast extract, 0.5 g NaCl, autoclaved,

then prior to use, 2 ml of 20 percent glucose and 1 ml of  $\text{MgCl}_2/\text{MgSO}_4$  (12g  $\text{MgSO}_4/9.5$  g  $\text{MgCl}_2/100$  ml water) per 100 ml of solution were added and filter sterilized] were added, and the mixture was incubated for one hour at 37°C with shaking at 225 rpm.

#### D. Screening of E. coli Colonies

Transformed cells carrying the correct insertion were identified by plating transformation mixture on appropriate antibiotic plates. Thus, 15 amp-resistant colonies (plated on media containing 50  $\mu\text{g/ml}$  of ampicillin) were picked. The inserted DNA was isolated using the minilysate method [Ish-Horowicz et al., Nucl. Acids Res., 9:2989 (1981)] and a portion of the DNA was cut with BamHI and PstI to generate a 483 bp fragment. The DNA samples were analyzed on 1 percent ethidium bromide-stained agarose gels to confirm the correct clones with the 483 bp DNA band. One of the plasmids so formed, designated plasmid "127", was induced with IPTG (isopropylthiogalactosidase) to give the targeted  $\beta$ -gal fusion polypeptide which was subsequently reacted with MB47 in mapping studies.

#### E. MB47 Binding Studies

The binding properties of monoclonal antibody MB47 with the expressed apo B-100 polypeptide fragments were then determined. Briefly, SURE E. coli cells expressing the fusion polypeptide encoded by the "127" plasmid or other plasmids were harvested. Cell extracts were clarified and their protein concentration was determined by a modified Lowry assay [Markwell et al., Anal. Biochem., 87:206 (1978)].

Proteinaceous bands were separated by electrophoresis of the extracts on polyacrylamide

gradient (3-20 percent) gels containing 1 percent SDS (sodium dodecylsulfate). The bands in the gel were blotted onto Immune-Lite blotting membrane (BioRad; Richmond, CA) using a semi-dry blotter. The presence  
5 of the desired expression product in the isolated bands was confirmed with monoclonal antibodies AI-11 and MB47 or for  $\beta$ -galactosidase activity using the Immune-Lite chemiluminescent kit (BioRad).

Fusion polypeptide-containing inclusion bodies  
10 were prepared according to the protocol of Pease et al., J. Biol. Chem., 265:553-568 (1990). Briefly, competent E. coli cells were transformed with the "127" plasmid or other plasmids encoding a fusion polypeptide of interest as described above. The  
15 transformed cells were grown in a rich nutrient broth (TYE broth) supplemented with 50  $\mu$ g/ml ampicillin, and production of the fusion polypeptide was induced with IPTG. E. coli cells containing the fusion polypeptide were collected by centrifugation and the cells were  
20 lysed by the addition of lysozyme to a concentration of 0.2 mg/ml.

The lysed cells were then treated with bovine pancreatic deoxyribonuclease and sodium deoxycholate. This admixture was subjected to high speed  
25 centrifugation to pellet the fusion protein-containing inclusion bodies. This pellet was resuspended in a solution of 50 mM Tris-HCl, 75 mM NaCl and 1 mM EDTA (Solution A, pH 7.5) containing sodium deoxycholate. This admixture was again centrifuged at high speed,  
30 after which the pellet was resuspended in Solution A plus 2-mercaptoethanol (70 mM final concentration) and Triton-X100 (1 percent final concentration). After a 30 minute incubation, this admixture was subjected to high speed centrifugation.

In a departure from the method of Pease et al., the fusion polypeptides were purified from the inclusion bodies by the following protocol.

Contaminating proteins were precipitated by the addition of 20 percent saturated ammonium sulfate to the supernatant fraction. These proteins were removed by high speed centrifugation, and the supernatant fraction from this step was also treated with 20 percent saturated ammonium sulfate. After another high speed centrifugation step, the supernatant fraction containing the fusion polypeptides was collected and dialyzed overnight in phosphate-buffered saline. After dialysis, the solution was clarified by brief high speed centrifugation. This procedure provided a dispersible fusion polypeptide substantially free of a denaturing surfactant in that the Triton X-100 was substantially removed during purification, and SDS was never used.

A Dynatech Immulon 2 microtiter plate was coated with the purified fusion polypeptide obtained above. The fusion polypeptide samples were diluted to a concentration of about 45  $\mu\text{g}$  using 10 mM sodium carbonate-bicarbonate buffer solution (pH 9.6). Wells in the microtiter plate were coated with 100  $\mu\text{l}$  of the diluted fusion polypeptide solution. The plate was incubated for about 18 hours at 4°C covered with an acetate plate cover, after which the plate was washed once with a standard PBS (phosphate buffered saline)-Tween wash solution (0.05 percent Tween 20, 0.05 M phosphate buffer, pH = 7.6). Nonspecific binding sites of the proteins were blocked with 200  $\mu\text{l}$  of 3 percent BSA (bovine serum albumin) in PBS and the admixture was incubated at 37°C for one hour.

A labeled MB47 solution was prepared using MB47 conjugated to HRP0 (horseradish peroxidase) as

described by P.K. Nakane, Immunofluorescence and Related Techniques, W. Knapp ed., (1978) Elsevier, Holland, pp.215. The HRP-conjugated MB47 was diluted with the 3 percent BSA in PBS solution to give a 0.1 percent conjugate solution.

Approximately 100  $\mu$ l of the conjugate solution was added to the fusion polypeptide-containing wells, covered with an acetate plate cover, and incubated at room temperature on an orbital shaker for one hour. The plate was then washed five times with the standard PBS/Tween solution.

Approximately 100  $\mu$ l of an o-phenylenediamine (OPD; Sigma; St. Louis, MO) substrate solution was added to each well and the admixture was incubated at room temperature for 30 minutes. The reaction was quenched with 50  $\mu$ l of 4N H<sub>2</sub>SO<sub>4</sub> per well and the plate was read at 490 nm. Notably, the measurements of optical density at 490 nm, relative to a control polypeptide, were high enough to indicate MB47 reactivity without a need for or hinderance of protein solubilization by SDS.

The results of these studies indicated that the affinities of MB47 for the examined fusion polypeptides can be classified into at least three groups. In the first group of polypeptides, MB47 immunoreacted strongly with the polypeptide fragments encoded by the about 483, 408, 345, and 282 base pair sequences corresponding to polypeptides s5-s8 of Table 2. In fact, the immunoaffinity of MB47 for these apo B-100 fragments is comparable to that for the about 1128 bp, 377 residue apo B-100 fragment (s9 in Table 1).

The above studies also revealed that the 471, 372, 276 and 243 bp cDNA noted in Table 2 code for apo B-100 polypeptide fragments s1-s4 also immunoreacted

strongly with MB47. However, the affinity of this second group of fusion polypeptides for MB47 is notably less than that for the first group of proteins. Finally, a third group of fusion polypeptides, encoded by the 171, 111, 90 and 45 bp nucleotide sequences noted in Table 2, showed little affinity for MB47.

Example 2: Preparation of apo A-I/B-100 Dual Constructs

Apo A-I/B-100 fusion proteins were prepared as further fusions with a  $\beta$ -galactosidase fragment. The syntheses are exemplified by the procedure used to form plasmid "137", which is described below.

A. Amplification of Apo A-I cDNA

A 663 bp nucleotide sequence of apo A-I cDNA was PCR-amplified from plasmids containing the cDNA. Briefly, human apo A-I cDNA containing a 696 bp sequence including the desired sequence was obtained from Dr. Jan Breslow of the Rockefeller University (New York, NY). The cDNA contained PstI restriction sites at either end of the sequence. The cDNA was cloned into the pBR322 vector (Pharmacia; Piscataway, NJ) and subjected to PstI restriction under standard conditions. Vector pBR322 has a PstI restriction site within the amp gene.

After ligation and transformation, the resulting transformed hosts were screened for ampicillin-sensitivity and tetracycline resistance. A plasmid derived from one such ampicillin-sensitive, tetracycline-resistant transformed host was designated plasmid pA101.

Approximately 5  $\mu$ g of pA101 was digested with 50 units of Pst I restriction enzyme and the

linearized DNA was precipitated with ethanol. Approximately 100 ng of collected DNA was used as a PCR template (Perkin Elmer) to amplify a 663 bp apo A-1 cDNA that did not contain the signal sequence (residues 1-18). The forward primer, designated oligonucleotide #90, was prepared by Research Genetics (Huntsville, AL) and had the following nucleotide sequence:

5' - ATGTCTGCAGCGGCATTTCTGGCAGCAA - 3'  
PstI (SEQ ID NO:19)

The reverse primer, designated oligonucleotide #72, was prepared by Genetic Design (Houston, TX) and had the following nucleotide sequence:

5' - GTGCCCTGCAGTTATCATTGGCGGAGGTCCTCGAGCGC - 3'  
PstI (SEQ ID NO:20)

B. Cloning and Identification of Apo A-I

The PCR generated apo A-I product was digested with 20 units of PstI restriction enzyme and ligated into the PstI site of the plasmid pUR291 (Pharmacia). The resultant plasmids were screened for proper insertion. SURE E. coli (Stratagene Cloning Systems; La Jolla, CA) was transformed with the plasmid and 100 amp-resistant colonies (plated on media containing 50 µg/ml of ampicillin) were picked on a grid and screened by hybridization with oligonucleotide primer #90 radioactively labeled with <sup>32</sup>P. Ten positive colonies were selected and checked for correct orientation by digestion with BamHI and XhoI restriction enzymes. Plasmids with the proper orientation generated the predicted 1200 bp fragment. One of the plasmids, designated plasmid "85", was used in formation of the apo A-I/B-100 dual construct of plasmid "137" as described below.



C. Formation of apo A-I/B-100 Dual Construct

Synthesis of apo A-I/B-100 dual construct plasmid "137" from apo A-I plasmid "85" was performed as described below. Briefly, plasmid "85" was  
5 linearized by digestion with HindIII at a unique site within the polylinker of original plasmid pUR291. The cohesive termini on the linearized DNA were converted to blunt-ends by digestion with T4 polymerase according to the manufacturer's instructions.

10 The linearized plasmid was cleaved at the unique XhoI site present at the 3' terminus of the apo A-I gene. The resultant linearized fragment of plasmid "85" had a cohesive XhoI site at the 3' terminus of the apo A-I gene and a blunt end at the 5'  
15 end of the plasmid.

A 483 bp apo B-100 insert was generated by PCR. In particular, plasmid "1.5Kb" (Pease et al., supra), which contained a 1.5kb apo B-100 insert in pUR291 was used as a template in a PCR protocol to amplify the  
20 483 bp fragment domain. This region was previously identified (in plasmid "127") as containing all of the information necessary for optimal MB47 binding. The 483 bp region was amplified using forward and reverse primers #132 and #130 (as shown in Table 3). A 483 bp  
25 PCR product having an XhoI site upstream of the 5' sequence terminus and a PstI site downstream of the 3' sequence terminus was thereby prepared, which also contained a 5 bp "buffer" sequence at the 3' terminus. Digestion of the PCR-amplified DNA with XhoI thereby  
30 generated a cohesive XhoI 5' terminus and a blunt 3' end.

The PCR-generated apo B-100 DNA fragment from plasmid "1.5Kb" obtained above was ligated directly into the before-discussed linearized plasmid "85"  
35 having a blunt 5' end and a XhoI site at the 3' end

using the protocol outlined by the vendor (Pharmacia). Plasmids containing the apo A-I/B-100 insert were screened for the correct placement and orientation by restriction endonuclease double digests using  
5 BamHI/XhoI and XhoI/PstI. A correct clone, designated plasmid "137", was identified and used in subsequent transformations of SURE E. coli. Expressed fusion polypeptides were analyzed on Western blots to confirm the presence of the dual epitope following a procedure  
10 like that described above for apo B-100 polypeptides.

Example 3: Assay of Plasma with Apo A-I/B-100  
Standard

An assay of human plasma for its LDL and HDL  
15 content can be performed using an above-described apo A-I/B-100 fusion protein as a reference standard. The feasibility of such a reference standard was demonstrated by the following study.

20 A. Isolation of Dual Construct and Control Proteins

Cells containing the "137" or pUR291 (control) plasmids were fermented at 37°C for 9 hours (including induction) to obtain a cell paste. The cell pastes  
25 were resuspended in sonication buffer (10 mg/g cells). The sonication buffer was prepared from Buffer A (0.1 M Tris-HCl, pH=7.8; 1 mM MgCl<sub>2</sub>; 0.5 mM Na<sub>2</sub>EDTA; 0.1 M NaCl) containing 0.1 M L-arginine hydrochloride, 10 mg/l PMSF (phenyl methyl sulfonyl fluoride), and 70  
30 mM 2-ME (2-mercaptoethanol) freshly added. Lysozyme was added to a final concentration of 0.5 mg/ml and the mixture was maintained for 20 minutes. Cells were disrupted by sonication (Artek; Farmingdale, NY) at 80 watts for five minutes on ice.

5 The raw extract was centrifuged at 10000 x g  
for 10 minutes, the supernatant discarded, and the  
pellet was resuspended in the same volume of Buffer A  
containing 70 mM 2-ME. The cell paste obtained was  
treated as previously described. After sonication and  
subsequent centrifugation, the supernatant was  
discarded and the pellet was resuspended in PBS  
containing 5 percent SDS buffer (and 70 mM 2-ME) at  
2.5 ml/g cells to give a stock solution of the  
10 protein.

B. Immunoaffinity Studies on Dual Construct  
Versus Control Proteins

15 A Dynatech-Immulon 2 microtiter plate was  
coated with varying dilutions of the above stock apo  
A-I/B-100 fusion protein and control proteins. A  
stock solution containing 0.95 µg/100 µl of the  
rehydrated apo A-I/B-100 fusion protein expressed by  
plasmid "137" was diluted at 1/2, 1/4, 1/8, 1/16,  
20 1/32, and 1/64 dilutions using a 10 mM sodium  
carbonate-bicarbonate buffer (pH 9.0). A pUR291 stock  
solution (prepared at 395.0 µg/100 µl) was similarly  
diluted. The microtiter wells were each coated with  
approximately 100 µl of solution - two wells for each  
25 protein at each dilution. Each fusion polypeptide was  
assayed with MB47 and AI-11 antibodies.

The coated microtiter plate was covered with an  
acetate plate and incubated about 18 hours (overnight)  
at 4°C. The plate was washed with the above standard  
30 PBS-Tween wash buffer and was blocked with 200 µl of a  
PBS + 3 percent BSA solution and was incubated at 37°C  
for one hour.

A 1:1000 dilution of HRPO-labeled MB47  
antibodies was prepared using the PBS + 3 percent BSA

diluent. A 1:1000 dilution of HRP0-labeled AI-11 was prepared using the PBS + 3 percent BSA diluent.

Approximately 100  $\mu$ l of MB47 conjugate were added to two sets of wells and approximately 100  $\mu$ l of the AI-11 conjugate were added to the other two sets of wells. The wells were covered with an acetate plate and incubated at room temperature on an orbital shaker for one hour. The plate was washed five times with PBS/Tween. Approximately 100  $\mu$ l of an OPD substrate were added to each well and the solutions were incubated for 30 minutes at room temperature. The reactions were then quenched with 4N H<sub>2</sub>SO<sub>4</sub> and read at 490 nm. The results are presented in Table 5 below.

TABLE 5

ELISA of Dual Construct ("137") Fusion Protein and Control (pUR291) Binding to MB47 and AI-11

Dilution <sup>2</sup>	MB47 <sup>1</sup>		AI-11 <sup>1</sup>	
	"137"	pUR291	"137"	pUR291
1/2	1.3	0.10	1.0	0.10
1/4	0.46	0.12	0.60	0.08
1/8	0.14	0.10	0.20	0.08
1/16	0.10	0.04	0.14	0.07
1/32	0.10	0.02	0.14	0.05
1/64	0.08	0.01	0.12	0.04

<sup>1</sup> Optical density at 490 nm.

<sup>2</sup> Dilution of stock solutions

The results of the studies described above indicated preferential binding between the assayed antibodies and the dispersible apo A-I/B-100 fusion polypeptide. Significantly, the results demonstrate

that the fusion polypeptide does not need to be solubilized, e.g., in a denaturing concentration of SDS (sodium dodecyl sulfate), for use.

C. Studies in Urea-Free Buffers

5 A cell preparation as discussed in (A), above, was again used as the source of the fusion polypeptide. Here, however, the buffer system used contained PBS and 5 percent SDS to solubilize the E.coli lysate following sonication.

10 After three sonications and resuspension into the buffer after a 30 minute, 16,000Xg centrifugation, the fusion stock solution prepared from the supernatant was loaded onto a model 491 Bio-Rad preparative SDS-polyacrylamide gel electrophoresis system. Two ml fractions were collected and dialyzed  
15 with 25 mM ammonium bicarbonate to remove substantially all of the SDS.

Fifty  $\mu$ l aliquots from every fifth dialyzed fraction were coated on the walls of 96-well  
20 polystyrene microtiter plates, incubated for one hour at room temperature and washed as discussed in (B), above, and then non-specific binding sites were blocked with BSA in PBS as noted in (B), above. Fifty  $\mu$ l of purified monoclonal antibodies AI-11 and MB47 at  
25 2.17 mg/ml and 2.44 mg/ml in 3 percent BSA in PBS were separately admixed with the coated wells and maintained at 4°C for about 18 hours (overnight). After washing the plates to remove unreacted antibody, radiolabeled ( $^{125}$ I) goat anti-mouse IgG (at  $2.2 \times 10^5$   
30 cpm/ $\mu$ l) (50  $\mu$ l in 97.2 percent TCA) was added to each well and maintained at 4°C for four hours. After washing, the bound radioactivity was counted.

The results of this study indicated that proteins with AI-11 and MB47 co-eluted from the  
35 preparative gel. Although a minimal amount of SDS was

likely present in the fusion polypeptide preparation, that amount did not inhibit use of the fusion polypeptide as an antigen for these studies, and that fusion polypeptide was substantially free of the denaturing surfactant.

Twenty-five fusion polypeptide-containing fractions (50 ml) were pooled and studied in another, similar study that also included HDL and LDL as coatings on the walls of the plates. The results of this study showed that there was sufficient fusion polypeptide in the pool, without further concentration, to perform HDL and LDL assays with monoclonals AI-11 and MB47, respectively. Those results also again showed that the minimal amount of SDS present in the fusion polypeptide preparation did not interfere with the antigenicity of the A-I/B-100 fusion polypeptide as is usually the case. Although an ammonium bicarbonate buffer was used for these studies, other buffers normally used for coating microtiter plates can also be used.

Although the present invention has been described in some detail by way of illustration and example for purposes of clarity and understanding, certain obvious modifications can be practiced within the scope of the appended claims.

## SEQUENCE LISTING

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- (iii) NUMBER OF SEQUENCES: 20
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  - (C) OPERATING SYSTEM: PC-DOS/MS-DOS
  - (D) SOFTWARE: PatentIn Release #1.0, Version #1.25
- (vi) CURRENT APPLICATION DATA:
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- (vii) PRIOR APPLICATION DATA:
  - (A) APPLICATION NUMBER: US 07/901,706
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## (2) INFORMATION FOR SEQ ID NO:1:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 377 amino acids
  - (B) TYPE: amino acid
  - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:1:  

Glu	Leu	Pro	Arg	Thr	Phe	Gln	Ile	Pro	Gly	Tyr	Thr	Val	Pro	Val	Val	
1				5					10					15		
Asn	Val	Glu	Val	Ser	Pro	Phe	Thr	Ile	Glu	Met	Ser	Ala	Phe	Gly	Tyr	
		20						25					30			
Val	Phe	Pro	Lys	Ala	Val	Ser	Met	Pro	Ser	Phe	Ser	Ile	Leu	Gly	Ser	
		35					40						45			

Asp Val Arg Val Pro Ser Tyr Thr Leu Ile Leu Pro Ser Leu Glu Leu  
 50 55 60  
 Pro Val Leu His Val Pro Arg Asn Leu Lys Leu Ser Leu Pro Asp Phe  
 65 70 75 80  
 Lys Glu Leu Cys Thr Ile Ser His Ile Phe Ile Pro Ala Met Gly Asn  
 85 90 95  
 Ile Thr Tyr Asp Phe Ser Phe Lys Ser Ser Val Ile Thr Leu Asn Thr  
 100 105 110  
 Asn Ala Glu Leu Phe Asn Gln Ser Asp Ile Val Ala His Leu Leu Ser  
 115 120 125  
 Ser Ser Ser Ser Val Ile Asp Ala Leu Gln Tyr Lys Leu Glu Gly Thr  
 130 135 140  
 Thr Arg Leu Thr Arg Lys Arg Gly Leu Lys Leu Ala Thr Ala Leu Ser  
 145 150 155 160  
 Leu Ser Asn Lys Phe Val Glu Gly Ser His Asn Ser Thr Val Ser Leu  
 165 170 175  
 Thr Thr Lys Asn Met Glu Val Ser Val Ala Thr Thr Thr Lys Ala Gln  
 180 185 190  
 Ile Pro Ile Leu Arg Met Asn Phe Lys Gln Glu Leu Asn Gly Asn Thr  
 195 200 205  
 Lys Ser Lys Pro Thr Val Ser Ser Ser Met Glu Phe Lys Tyr Asp Phe  
 210 215 220  
 Asn Ser Ser Met Leu Tyr Ser Thr Ala Lys Gly Ala Val Asp His Lys  
 225 230 235 240  
 Leu Ser Leu Glu Ser Leu Thr Ser Tyr Phe Ser Ile Glu Ser Ser Thr  
 245 250 255  
 Lys Gly Asp Val Lys Gly Ser Val Leu Ser Arg Glu Tyr Ser Gly Thr  
 260 265 270  
 Ile Ala Ser Glu Ala Asn Thr Tyr Leu Asn Ser Lys Ser Thr Arg Ser  
 275 280 285  
 Ser Val Lys Leu Gln Gly Thr Ser Lys Ile Asp Asp Ile Trp Asn Leu  
 290 295 300  
 Glu Val Lys Glu Asn Phe Ala Gly Glu Ala Thr Leu Gln Arg Ile Tyr  
 305 310 315 320  
 Ser Leu Trp Glu His Ser Thr Lys Asn His Leu Gln Leu Glu Gly Leu  
 325 330 335  
 Phe Phe Thr Asn Gly Glu His Thr Ser Lys Ala Thr Leu Glu Leu Ser  
 340 345 350  
 Pro Trp Gln Met Ser Ala Leu Val Gln Val His Ala Ser Gln Pro Ser  
 355 360 365  
 Ser Phe His Asp Phe Pro Asp Leu Gly  
 370 375



## (2) INFORMATION FOR SEQ ID NO:2:

## (i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 1131 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

## (ii) MOLECULE TYPE: DNA (genomic)

## (ix) FEATURE:

- (A) NAME/KEY: CDS
- (B) LOCATION: 1..1131

## (xi) SEQUENCE DESCRIPTION: SEQ ID NO:2:

GAG CTC CCC AGG ACC TTT CAA ATT CCT GGA TAC ACT GTT CCA GTT GTC Glu Leu Pro Arg Thr Phe Gln Ile Pro Gly Tyr Thr Val Pro Val Val 1 5 10 15	48
AAT GTT GAA GTG TCT CCA TTC ACC ATA GAG ATG TCG GCA TTC GGC TAT Asn Val Glu Val Ser Pro Phe Thr Ile Glu Met Ser Ala Phe Gly Tyr 20 25 30	96
GTG TTC CCA AAA GCA GTC AGC ATG CCT AGT TTC TCC ATC CTA GGT TCT Val Phe Pro Lys Ala Val Ser Met Pro Ser Phe Ser Ile Leu Gly Ser 35 40 45	144
GAC GTC CGT GTG CCT TCA TAC ACA TTA ATC CTG CCA TCA TTA GAG CTG Asp Val Arg Val Pro Ser Tyr Thr Leu Ile Leu Pro Ser Leu Glu Leu 50 55 60	192
CCA GTC CTT CAT GTC CCT AGA AAT CTC AAG CTT TCT CTT CCA GAT TTC Pro Val Leu His Val Pro Arg Asn Leu Lys Leu Ser Leu Pro Asp Phe 65 70 75 80	240
AAG GAA TTG TGT ACC ATA AGC CAT ATT TTT ATT CCT GCC ATG GGC AAT Lys Glu Leu Cys Thr Ile Ser His Ile Phe Ile Pro Ala Met Gly Asn 85 90 95	288
ATT ACC TAT GAT TTC TCC TTT AAA TCA AGT GTC ATC ACA CTG AAT ACC Ile Thr Tyr Asp Phe Ser Phe Lys Ser Ser Val Ile Thr Leu Asn Thr 100 105 110	336
AAT GCT GAA CTT TTT AAC CAG TCA GAT ATT GTT GCT CAT CTC CTT TCT Asn Ala Glu Leu Phe Asn Gln Ser Asp Ile Val Ala His Leu Leu Ser 115 120 125	384
TCA TCT TCA TCT GTC ATT GAT GCA CTG CAG TAC AAA TTA GAG GGC ACC Ser Ser Ser Ser Val Ile Asp Ala Leu Gln Tyr Lys Leu Glu Gly Thr 130 135 140	432
ACA AGA TTG ACA AGA AAA AGG GGA TTG AAG TTA GCC ACA GCT CTG TCT Thr Arg Leu Thr Arg Lys Arg Gly Leu Lys Leu Ala Thr Ala Leu Ser 145 150 155 160	480
CTG AGC AAC AAA TTT GTG GAG GGT AGT CAT AAC AGT ACT GTG AGC TTA Leu Ser Asn Lys Phe Val Glu Gly Ser His Asn Ser Thr Val Ser Leu 165 170 175	528
ACC ACG AAA AAT ATG GAA GTG TCA GTG GCA ACA ACC ACA AAA GCC CAA Thr Thr Lys Asn Met Glu Val Ser Val Ala Thr Thr Thr Lys Ala Gln 180 185 190	576
ATT CCA ATT TTG AGA ATG AAT TTC AAG CAA GAA CTT AAT GGA AAT ACC Ile Pro Ile Leu Arg Met Asn Phe Lys Gln Glu Leu Asn Gly Asn Thr 195 200 205	624

AAG TCA AAA CCT ACT GTC TCT TCC TCC ATG GAA TTT AAG TAT GAT TTC Lys Ser Lys Pro Thr Val Ser Ser Ser Met Glu Phe Lys Tyr Asp Phe 210 215 220	672
AAT TCT TCA ATG CTG TAC TCT ACC GCT AAA GGA GCA GTT GAC CAC AAG Asn Ser Ser Met Leu Tyr Ser Thr Ala Lys Gly Ala Val Asp His Lys 225 230 235 240	720
CTT AGC TTG GAA AGC CTC ACC TCT TAC TTT TCC ATT GAG TCA TCT ACC Leu Ser Leu Glu Ser Leu Thr Ser Tyr Phe Ser Ile Glu Ser Ser Thr 245 250 255	768
AAA GGA GAT GTC AAG GGT TCG GTT CTT TCT CGG GAA TAT TCA GGA ACT Lys Gly Asp Val Lys Gly Ser Val Leu Ser Arg Glu Tyr Ser Gly Thr 260 265 270	816
ATT GCT AGT GAG GCC AAC ACT TAC TTG AAT TCC AAG AGC ACA CGG TCT Ile Ala Ser Glu Ala Asn Thr Tyr Leu Asn Ser Lys Ser Thr Arg Ser 275 280 285	864
TCA GTG AAG CTG CAG GGC ACT TCC AAA ATT GAT GAT ATC TGG AAC CTT Ser Val Lys Leu Gln Gly Thr Ser Lys Ile Asp Asp Ile Trp Asn Leu 290 295 300	912
GAA GTA AAA GAA AAT TTT GCT GGA GAA GCC ACA CTC CAA CGC ATA TAT Glu Val Lys Glu Asn Phe Ala Gly Glu Ala Thr Leu Gln Arg Ile Tyr 305 310 315 320	960
TCC CTC TGG GAG CAC AGT ACG AAA AAC CAC TTA CAG CTA GAG GGC CTC Ser Leu Trp Glu His Ser Thr Lys Asn His Leu Gln Leu Glu Gly Leu 325 330 335	1008
TTT TTC ACC AAC GGA GAA CAT ACA AGC AAA GCC ACC CTG GAA CTC TCT Phe Phe Thr Asn Gly Glu His Thr Ser Lys Ala Thr Leu Glu Leu Ser 340 345 350	1056
CCA TGG CAA ATG TCA GCT CTT GTT CAG GTC CAT GCA AGT CAG CCC AGT Pro Trp Gln Met Ser Ala Leu Val Gln Val His Ala Ser Gln Pro Ser 355 360 365	1104
TCC TTC CAT GAT TTC CCT GAC CTT GGC Ser Phe His Asp Phe Pro Asp Leu Gly 370 375	1131

## (2) INFORMATION FOR SEQ ID NO:3:

## (i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 267 amino acids
- (B) TYPE: amino acid
- (D) TOPOLOGY: linear

## (ii) MOLECULE TYPE: protein

## (xi) SEQUENCE DESCRIPTION: SEQ ID NO:3:

Met Lys Ala Ala Val Leu Thr Leu Ala Val Leu Phe Leu Thr Gly Ser 1 5 10 15
Gln Ala Arg His Phe Trp Gln Gln Asp Glu Pro Pro Gln Ser Pro Trp 20 25 30
Asp Arg Val Lys Asp Leu Ala Thr Val Tyr Val Asp Val Leu Lys Asp 35 40 45

Ser Gly Arg Asp Tyr Val Ser Gln Phe Glu Gly Ser Ala Leu Gly Lys  
 50 55 60  
 Gln Leu Asn Leu Lys Leu Leu Asp Asn Trp Asp Ser Val Thr Ser Thr  
 65 70 75 80  
 Phe Ser Lys Leu Arg Glu Gln Leu Gly Pro Val Thr Gln Glu Phe Trp  
 85 90 95  
 Asp Asn Leu Glu Lys Glu Thr Glu Gly Leu Arg Gln Glu Met Ser Lys  
 100 105 110  
 Asp Leu Glu Glu Val Lys Ala Lys Val Gln Pro Tyr Leu Asp Asp Phe  
 115 120 125  
 Gln Lys Lys Trp Gln Glu Glu Met Glu Leu Tyr Arg Gln Lys Val Glu  
 130 135 140  
 Pro Leu Arg Ala Glu Leu Gln Glu Gly Ala Arg Gln Lys Leu His Glu  
 145 150 155 160  
 Leu Gln Glu Lys Leu Ser Pro Leu Gly Glu Glu Met Arg Asp Arg Ala  
 165 170 175  
 Arg Ala His Val Asp Ala Leu Arg Thr His Leu Ala Pro Tyr Ser Asp  
 180 185 190  
 Glu Leu Arg Gln Arg Leu Ala Ala Arg Leu Glu Ala Leu Lys Glu Asn  
 195 200 205  
 Gly Gly Ala Arg Leu Ala Glu Tyr His Ala Lys Ala Thr Glu His Leu  
 210 215 220  
 Ser Thr Leu Ser Glu Lys Ala Lys Pro Ala Leu Glu Asp Leu Arg Gln  
 225 230 235 240  
 Gly Leu Leu Pro Val Leu Glu Ser Phe Lys Val Ser Phe Leu Ser Ala  
 245 250 255  
 Leu Glu Glu Tyr Thr Lys Lys Leu Asn Thr Gln  
 260 265

## (2) INFORMATION FOR SEQ ID NO:4:

## (i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 801 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

## (ii) MOLECULE TYPE: DNA (genomic)

## (ix) FEATURE:

- (A) NAME/KEY: CDS
- (B) LOCATION: 1..801

## (xi) SEQUENCE DESCRIPTION: SEQ ID NO:4:

ATG AAA GCT GCC GTG CTG ACC TTG GCC GTG CTC TTC CTG ACG GGG AGC	48
Met Lys Ala Ala Val Leu Thr Leu Ala Val Leu Phe Leu Thr Gly Ser	
1 5 10 15	
CAG GCT CGG CAT TTC TGG CAG CAA GAT GAA CCC CCC CAG AGC CCC TGG	96
Gln Ala Arg His Phe Trp Gln Gln Asp Glu Pro Pro Gln Ser Pro Trp	
20 25 30	

GAT CGA GTG AAG GAC CTG GCC ACT GTG TAC GTG GAT GTG CTC AAA GAC Asp Arg Val Lys Asp Leu Ala Thr Val Tyr Val Asp Val Leu Lys Asp 35 40 45	144
AGC GGC AGA GAC TAT GTG TCC CAG TTT GAA GGC TCC GCC TTG GGA AAA Ser Gly Arg Asp Tyr Val Ser Gln Phe Glu Gly Ser Ala Leu Gly Lys 50 55 60	192
CAG CTA AAC CTA AAG CTC CTT GAC AAC TGG GAC AGC GTG ACC TCC ACC Gln Leu Asn Leu Lys Leu Leu Asp Asn Trp Asp Ser Val Thr Ser Thr 65 70 75 80	240
TTC AGC AAG CTG CGC GAA CAG CTC GGC CCT GTG ACC CAG GAG TTC TGG Phe Ser Lys Leu Arg Glu Gln Leu Gly Pro Val Thr Gln Glu Phe Trp 85 90 95	288
GAT AAC CTG GAA AAG GAG ACA GAG GGC CTG AGG CAG GAG ATG AGC AAG Asp Asn Leu Glu Lys Glu Thr Glu Gly Leu Arg Gln Glu Met Ser Lys 100 105 110	336
GAT CTG GAG GAG GTG AAG GCC AAG GTG CAG CCC TAC CTG GAC GAC TTC Asp Leu Glu Glu Val Lys Ala Lys Val Gln Pro Tyr Leu Asp Asp Phe 115 120 125	384
CAG AAG AAG TGG CAG GAG GAG ATG GAG CTC TAC CGC CAG AAG GTG GAG Gln Lys Lys Trp Gln Glu Glu Met Glu Leu Tyr Arg Gln Lys Val Glu 130 135 140	432
CCG CTG CGC GCA GAG CTC CAA GAG GGC GCG CGC CAG AAG CTG CAC GAG Pro Leu Arg Ala Glu Leu Gln Glu Gly Ala Arg Gln Lys Leu His Glu 145 150 155 160	480
CTG CAA GAG AAG CTG AGC CCA CTG GGC GAG GAG ATG CGC GAC CGC GCG Leu Gln Glu Lys Leu Ser Pro Leu Gly Glu Glu Met Arg Asp Arg Ala 165 170 175	528
CGC GCC CAT GTG GAC GCG CTG CGC ACG CAT CTG GCC CCC TAC AGC GAC Arg Ala His Val Asp Ala Leu Arg Thr His Leu Ala Pro Tyr Ser Asp 180 185 190	576
GAG CTG CGC CAG CGC TTG GCC GCG CGC CTT GAG GCT CTC AAG GAG AAC Glu Leu Arg Gln Arg Leu Ala Ala Arg Leu Glu Ala Leu Lys Glu Asn 195 200 205	624
GGC GGC GCC AGA CTG GCC GAG TAC CAC GCC AAG GCC ACC GAG CAT CTG Gly Gly Ala Arg Leu Ala Glu Tyr His Ala Lys Ala Thr Glu His Leu 210 215 220	672
AGC ACG CTC AGC GAG AAG GCC AAG CCC GCG CTC GAG GAC CTC CGC CAA Ser Thr Leu Ser Glu Lys Ala Lys Pro Ala Leu Glu Asp Leu Arg Gln 225 230 235 240	720
GGC CTG CTG CCC GTG CTG GAG AGC TTC AAG GTC AGC TTC CTG AGC GCT Gly Leu Leu Pro Val Leu Glu Ser Phe Lys Val Ser Phe Leu Ser Ala 245 250 255	768
CTC GAG GAG TAC ACT AAG AAG CTC AAC ACC CAG Leu Glu Glu Tyr Thr Lys Lys Leu Asn Thr Gln 260 265	801

## (2) INFORMATION FOR SEQ ID NO:5:

- (i) SEQUENCE CHARACTERISTICS:  
(A) LENGTH: 16 amino acids

(B) TYPE: amino acid  
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:5:

Lys Val Gln Pro Tyr Leu Asp Asp Phe Gln Lys Lys Trp Gln Glu Glu  
1 5 10 15

(2) INFORMATION FOR SEQ ID NO:6:

(i) SEQUENCE CHARACTERISTICS:  
(A) LENGTH: 5 amino acids  
(B) TYPE: amino acid  
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:6:

Gly Gly Gly Gly Ser  
1 5

(2) INFORMATION FOR SEQ ID NO:7:

(i) SEQUENCE CHARACTERISTICS:  
(A) LENGTH: 31 base pairs  
(B) TYPE: nucleic acid  
(C) STRANDEDNESS: single  
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:7:

ATGCGGATCC AAATTAGAGG GCACCACAAG A

31

(2) INFORMATION FOR SEQ ID NO:8:

(i) SEQUENCE CHARACTERISTICS:  
(A) LENGTH: 31 base pairs  
(B) TYPE: nucleic acid  
(C) STRANDEDNESS: single  
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:8:

ATGCGGATCC ACTGTGAGCT TAACCACGAA A

31

(2) INFORMATION FOR SEQ ID NO:9:

(i) SEQUENCE CHARACTERISTICS:  
(A) LENGTH: 29 base pairs  
(B) TYPE: nucleic acid

(C) STRANDEDNESS: single  
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:9:

ATGCGGATCC AATGAATACC AAGTCAAAA

29

(2) INFORMATION FOR SEQ ID NO:10:

(i) SEQUENCE CHARACTERISTICS:  
(A) LENGTH: 34 base pairs  
(B) TYPE: nucleic acid  
(C) STRANDEDNESS: single  
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:10:

ATGCGGATCC TCCTCCATGG AATTAAAGTA TGAT

34

(2) INFORMATION FOR SEQ ID NO:11:

(i) SEQUENCE CHARACTERISTICS:  
(A) LENGTH: 34 base pairs  
(B) TYPE: nucleic acid  
(C) STRANDEDNESS: single  
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:11:

ATGCCTCGAG TCCTCCATGG AATTAAAGTA TGAT

34

(2) INFORMATION FOR SEQ ID NO:12:

(i) SEQUENCE CHARACTERISTICS:  
(A) LENGTH: 43 base pairs  
(B) TYPE: nucleic acid  
(C) STRANDEDNESS: single  
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:12:

ATAGCTGCAG TTTGGAAGTG CCCTGGAGCT TCACTGAAGA CCG

43

(2) INFORMATION FOR SEQ ID NO:13:

(i) SEQUENCE CHARACTERISTICS:  
(A) LENGTH: 37 base pairs  
(B) TYPE: nucleic acid  
(C) STRANDEDNESS: single  
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:13:

TTAACTGCAG CTATTAGCCA AGGTCAGGGA AATCATG

37

## (2) INFORMATION FOR SEQ ID NO:14:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 37 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear

- (ii) MOLECULE TYPE: DNA (genomic)

- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:14:

AATTCTGCAG TTATCAGCCG GGAGAGAGTT CCAGGGT

37

## (2) INFORMATION FOR SEQ ID NO:15:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 37 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear

- (ii) MOLECULE TYPE: DNA (genomic)

- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:15:

AATTCTGCAG TTATCAGTGT AAGTGGTTTT TCGTACT

37

## (2) INFORMATION FOR SEQ ID NO:16:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 37 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear

- (ii) MOLECULE TYPE: DNA (genomic)

- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:16:

AATTCTGCAG TTATCAAAAA TTTTCTTTTA CTTCAAG

37

## (2) INFORMATION FOR SEQ ID NO:17:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 31 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear

- (ii) MOLECULE TYPE: DNA (genomic)

- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:17:

ATGCGGATCC AAATTAGAGG GCACCACAAG A

31

## (2) INFORMATION FOR SEQ ID NO:18:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 38 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: single

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:18:

GTGCCCTGCA GTTATCACTT CACTGAAGAC CGTGTGCT

38

(2) INFORMATION FOR SEQ ID NO:19:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 28 base pairs

(B) TYPE: nucleic acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:19:

ATGTCTGCAG CGGCATTCT GGCAGCAA

28

(2) INFORMATION FOR SEQ ID NO:20:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 38 base pairs

(B) TYPE: nucleic acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:20:

GTGCCCTGCA GTTATCAT TG GCGGAGGTCC TCGAGCGC

38



## What is Claimed is:

1. A nonnatural polypeptide having an amino acid residue sequence up through about 375 residues in length that includes the amino acid residue sequence of apo B-100 shown in SEQ ID NO:1 from about residue 217 through about residue 297, which polypeptide immunoreacts with antibodies secreted by the hybridoma MB47 having ATCC Accession No. HB 8746, said polypeptide being dispersible in an aqueous medium in the substantial absence of a denaturing surfactant.
2. The polypeptide of claim 1 that includes the amino acid residue sequence of apo B-100 from residue 216 through residue 310.
3. The polypeptide of claim 2 that includes the amino acid residue sequence of apo B-100 from residue 216 through residue 331.
4. The polypeptide of claim 3 that includes the amino acid residue sequence of apo B-100 from residue 216 through residue 352.
5. The polypeptide of claim 4 that includes the amino acid residue sequence of apo B-100 from residue 216 through residue 377.
6. The polypeptide of claim 5 that includes the amino acid residue sequence of apo B-100 from residue 1 through residue 377.

7. The polypeptide of claim 1 that includes an amino acid residue sequence of apo B-100 that is selected from the group consisting of residues 205 to 297, 173 through 297, 140 through 297 and 216 through 310.

8. The polypeptide of claim 1 further having a beta-galactosidase moiety operatively linked to the amino-terminus thereof.

9. The polypeptide of claim 1, wherein said polypeptide includes the amino acid residue sequence of apo B-100 from residue position 216 through position 377 operatively linked to the C-terminus beta-galactosidase.

10. An isolated DNA segment comprising a nucleotide sequence encoding the amino acid residue sequence of apo B-100 shown in SEQ ID NO:1 from about residue 217 through about residue 297.

11. An isolated DNA segment as in claim 10, wherein the nucleotide sequence encodes the amino acid residue sequence of apo B-100 from about residue 217 through about residue 307.

12. An isolated DNA segment as in claim 11, wherein the nucleotide sequence encodes the amino acid residue sequence of apo B-100 from about residue 217 through about residue 377.

13. A recombinant DNA molecule that is replicable upon transfection in a suitable host cell, said recombinant DNA molecule comprising a DNA segment encoding an amino acid residue sequence of apo B-100 shown in SEQ ID NO:1 from about residue 217 through about residue 297.

14. A strain of SURE E. coli transformed with the recombinant DNA molecule of claim 13.

15. An isolated DNA segment comprising up through about 663 nucleotides that encodes an amino acid residue sequence of apo A-I shown in SEQ ID NO:3 including the sequence from residue 120 through residue 135.

16. The isolated DNA segment of claim 15, wherein the amino acid residue sequence of apo A-I includes the sequence from about residue 19 through about residue 240.

17. A vector operatively linked to a DNA segment comprising up through about 663 nucleotides that encodes an amino acid residue sequence of apo A-I shown in SEQ ID NO:3 from about residue 120 through about residue 135.

18. The vector of claim 17, wherein the DNA segment encodes the amino acid residue sequence of apo A-I shown in SEQ ID NO:3 from about residue 19 through about residue 240.

19. A fusion polypeptide that contains:

(a) a first amino acid residue sequence up through about 250 residues in length that includes the amino acid residue sequence of apo A-I shown in SEQ ID NO:3 from about residue 120 through about residue 135, said fusion polypeptide immunoreacting with anti-apo A-I antibodies secreted by a hybridoma selected from the group consisting of:

AI-4 (ATCC HB 8744),  
AI-7 (ATCC HB 8745),  
AI-9 (ATCC HB 8741),  
AI-10 (ATCC HB 9200),  
AI-11 (ATCC HB 9201),  
AI-12 (ATCC HB 9202),  
AI-13 (ATCC HB 9203),  
AI-14 (ATCC HB 9204), and  
AI-18 (ATCC HB 9570); and

(b) a second amino acid residue sequence up through about 375 residues in length that includes the amino acid residue sequence of apo B-100 shown in SEQ ID NO:1 from about residue 217 through about residue 297, said fusion polypeptide immunoreacting with antibodies secreted by the hybridoma MB47 (ATCC HB 8746);

said second amino acid residue sequence operatively linked to the carboxy-terminus of said first amino acid residue sequence.

20. The fusion polypeptide of claim 19, wherein the first and second amino acid residue sequences are linked by a third amino acid residue sequence up through about 120 residues in length.

21. The fusion polypeptide of claim 20, wherein the first amino acid residue sequence includes the amino acid residue sequence of apo A-I shown in SEQ ID NO:3 from residue 19 through residue 240.

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22. A vector replicable upon transfection in a suitable host cell, said vector comprising:

(a) a first DNA segment having a 5' and a 3' end and up through about 696 nucleotides in length that encodes an amino acid residue sequence including the apo A-I sequence shown in SEQ ID NO:3 from about residue 120 through about residue 135; and

(b) a second DNA segment have a 5' and a 3' end and up through about 1130 nucleotides in length that encodes an amino acid residue sequence including the apo B-100 sequence shown in SEQ ID NO:1 from about residue 217 through about residue 297;

the 5' end of said second DNA segment operatively ligated to the 3' end of said first DNA segment.

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23. The vector of claim 22, wherein the first and second DNA segments are operatively ligated by a third DNA segment up through about 360 nucleotides in length.

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24. A strain of SURE E. coli transformed with the vector of claim 22.

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25. A method for expressing a polypeptide in SURE E. coli, said polypeptide including an amino acid residue sequence of apo B-100 shown in SEQ ID NO:1

including the sequence from about residue 217 through about residue 297, said method comprising:

5 maintaining SURE E. coli transformed with an expression vector encoding said polypeptide under culture conditions for a time period sufficient for the transformed E. coli to express the polypeptide.

26. The method of claim 25 including the further step of recovering the expressed polypeptide.

10

27. A method for assaying the amount of native apo B-100 in a body fluid sample comprising:

15 (a) admixing a predetermined amount of a body fluid sample containing native apo B-100 in a liquid medium with predetermined amounts of a polypeptide affixed to a solid support and pan anti-apo B-100 antibodies to form a solid-liquid admixture, said pan anti-apo B-100 antibodies immunoreacting with the native apo B-100 of the sample and with the polypeptide, said polypeptide having an amino acid residue sequence up through about 375 residues in length that includes the amino acid residue sequence of the apo B-100 sequence shown in SEQ ID NO:1 from about residue 217 through about residue 297;

20

25 (b) maintaining the solid-liquid admixture under biological assay conditions for a time period sufficient for the pan anti-apo B-100 antibodies to immunoreact with the polypeptide to form a solid-phase polypeptide/anti-apo B-100 immunocomplex and for the pan anti-apo B-100 antibodies to immunoreact with the apo B-100 to form an apo B-100/anti-apo B-100 immunocomplex; and

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(c) determining the amount of solid-phase polypeptide/anti-apo B-100 immunocomplex formed, and thereby the amount of native apo B-100 in the body fluid sample.

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28. The method of claim 27, wherein the body fluid sample, polypeptide, and pan anti-apo B-100 antibodies are admixed substantially simultaneously.

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29. The method of claim 30, wherein the pan anti-apo B-100 antibodies are secreted by the hybridoma MB47 having ATCC Accession No. HB 8746.

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30. The method of claim 30, wherein the anti-apo B-100 antibodies are conjugated to a marker selected from the group consisting of radioactive, enzyme, biotin, fluorescent, and chemiluminescent labels.

20

31. A method for assaying the amount of native apo B-100 in a body fluid sample comprising:

(a) admixing in a liquid medium a predetermined amount of a body fluid sample containing native apo B-100 with predetermined amounts of a polypeptide and pan anti-apo B-100 antibodies to form an admixture, said pan anti-apo B-100 antibodies immunoreacting with the native apo B-100 of the sample and with the polypeptide, said polypeptide having an amino acid residue sequence up through about 375 residues in length that includes the amino acid residue sequence of the apo B-100 sequence shown in

30

SEQ ID NO:1 from about residue 217 through about residue 297;

5 (b) maintaining the admixture under biological assay conditions for a time period sufficient for the anti-apo B-100 antibodies to immunoreact with the polypeptide to form a polypeptide/anti-apo B-100 immunocomplex and for those antibodies to form an apo B-100/anti-apo B-100 immunocomplex; and

10 (c) determining the amount of polypeptide/anti-apo B-100 immunocomplex formed, and thereby the amount of native apo B-100 in the body fluid sample.

15 32. The method of claim 31, wherein the body fluid sample, polypeptide, and anti-apo B-100 antibodies are admixed substantially simultaneously.

20 33. The method of claim 31, wherein the polypeptide is conjugated to a marker selected from the group consisting of radioactive, enzyme, biotin, fluorescent, and chemiluminescent labels.

25 34. The method of claim 31, wherein the determination step comprises:

(i) admixing said polypeptide/anti-apo B-100 immunocomplex with second antibodies immunoreactive with said polypeptide to form a second admixture, said second antibodies operatively linked to a marker;

30 (ii) maintaining said second admixture under biological assay conditions for a period of time



sufficient for said second antibodies to immunoreact with said polypeptide/anti-apo B-100 immunocomplex to form a second antibody/polypeptide/anti-apo B-100 sandwich complex; and

- 5                   (iii) determining the amount of marker affixed to said sandwich complex, thereby determining the amount of polypeptide/anti-apo B-100 immunocomplex formed.

10                  35. The method of claim 34, wherein the marker is selected from the group consisting of radioactive, enzyme, biotin, fluorescent, and chemiluminescent labels.

15                  36. The method of claim 31, wherein the anti-apo B-100 antibodies are affixed to a solid support prior to the admixture of body fluid sample and polypeptide.

20                  37. The method of claim 31, wherein the anti-apo B-100 antibodies are secreted by the hybridoma MB47 having ATCC Accession No. HB 8746.

25                  38. A method for standardizing an assay for the amount of native apo B-100 in a body fluid sample comprising:

- 30                   (a) admixing first predetermined amounts of a body fluid sample containing native apo B-100 and pan anti-apo B-100 antibodies in a liquid medium to form a first admixture, said pan anti-apo B-100 antibodies immunoreacting with the native apo B-100 of the sample and with a polypeptide, said polypeptide

having an amino acid residue sequence up through about 375 residues in length that includes the amino acid residue sequence of the apo B-100 sequence shown in SEQ ID NO:1 from about residue 217 through about residue 297;

(b) maintaining the first admixture under biological assay conditions for a time period sufficient for the anti-apo B-100 antibodies to immunoreact with the native apo B-100 in the sample to form a native apo B-100/anti-apo B-100 immunocomplex;

(c) admixing second predetermined amounts of said pan anti-apo B-100 antibodies and the polypeptide of step (a) in a liquid medium to form a second admixture, said pan anti-apo B-100 antibodies immunoreacting with said polypeptide;

(d) maintaining the second admixture under biological assay conditions for a time period sufficient for the pan anti-apo B-100 antibodies to immunoreact with the polypeptide to form a polypeptide/anti-apo B-100 immunocomplex; and

(e) determining the amounts of native apo B-100/anti-apo B-100 and polypeptide/anti-apo B-100 immunocomplexes formed, and thereby the amount of native apo B-100 in the body fluid sample.

39. The method of claim 38, wherein the pan anti-apo B-100 antibodies are conjugated to a marker selected from the group consisting of radioactive, enzyme, biotin, fluorescent, and chemiluminescent labels.

40. The method of claim 38, wherein the pan anti-apo B-100 antibodies are affixed to a solid support, prior to admixture of the body fluid sample or polypeptide.

5

41. The method of claim 38, wherein the pan anti-apo B-100 antibodies are secreted by the hybridoma MB47 having ATCC Accession No. HB 8746.

10

42. The method of claim 38, wherein said determination step comprises:

(i) admixing said native apo B-100/anti-apo B-100 immunocomplex with second antibodies immunoreactive with said native apo B-100 to form a third admixture, said second antibodies operatively linked to a first marker;

(ii) maintaining said third admixture under biological assay conditions for a period of time sufficient for said second antibodies to immunoreact with said native apo B-100/anti-apo B-100 immunocomplex to form a second antibody/native apo B-100/anti-apo B-100 sandwich complex;

(iii) determining the amount of first marker affixed to said second antibody/native apo B-100/anti-apo B-100 sandwich complex, thereby determining the amount of native apo B-100/anti-apo B-100 immunocomplex formed;

(iv) admixing said polypeptide/anti-apo B-100 immunocomplex with third antibodies immunoreactive with said polypeptide or bound pan apo B-100 antibodies to form a fourth admixture, said

third antibodies operatively linked to a second marker;

5 (v) maintaining said fourth admixture under biological assay conditions for a period of time sufficient for said third antibodies to immunoreact with said polypeptide/anti-apo B-100 immunocomplex to form a third antibody/polypeptide/ anti-apo B-100 sandwich complex; and

10 (vi) determining the amount of second marker affixed to said third antibody/polypeptide/anti-apo B-100 sandwich complex, thereby determining the amount of polypeptide/anti-apo B-100 immunocomplex formed.

15 43. The method of claim 42, wherein the second antibody is an MB24 antibody secreted by the hybridoma having ATCC Accession No. HB 8742, said polypeptide is a fusion protein that includes the amino acid residue sequence of beta-galactosidase and the third antibody  
20 is an anti-beta-galactosidase antibody.

44. The method of claim 42, wherein the first and second markers are identical.

25 45. The method of claim 38 wherein said liquid medium of step (c) includes a delipidated body fluid sample.

30 46. The method of claim 45 wherein the amount of delipidated body fluid sample is the same as the amount of body fluid sample used in step (a).

47. A method for assaying the amounts of native apo B-100 and native apo A-I in a body fluid sample comprising:

5 (a) admixing a body fluid sample in a liquid medium with predetermined amounts of a fusion polypeptide affixed to a solid support, pan anti-apo B-100 antibodies, and pan anti-apo A-I antibodies to form a solid-liquid admixture, said pan anti-apo B-100 antibodies immunoreacting with the native apo B-100 of  
10 the sample and with the fusion polypeptide, said pan anti-apo A-I antibodies immunoreacting with the native apo A-I of the sample and with the fusion polypeptide, said fusion polypeptide comprising:

15 (i) a first amino acid residue sequence up through about 250 residues in length that includes the amino acid residue sequence of apo A-I shown in SEQ ID NO:3 from about residue 120 through about residue 135, said second amino acid residue  
20 sequence of said fusion polypeptide immunoreacting with anti-apo A-I antibodies secreted by a hybridoma selected from the group consisting of:

AI-4 (ATCC HB 8744),  
AI-7 (ATCC HB 8745),  
AI-9 (ATCC HB 8741),  
25 AI-10 (ATCC HB 9200),  
AI-11 (ATCC HB 9201),  
AI-12 (ATCC HB 9202),  
AI-13 (ATCC HB 9203),  
AI-14 (ATCC HB 9204), and  
30 AI-18 (ATCC HB 9570);

(ii) a second amino acid residue sequence up through about 375 residues in length that

includes the amino acid residue sequence of the apo B-100 sequence shown in SEQ ID NO:1 from about residue 217 through about residue 297, said fusion polypeptide immunoreacting with antibodies secreted by the hybridoma MB47 (ATCC HB 8746);

said second amino acid residue sequence operatively linked to the carboxy terminus of said first amino acid residue sequence;

- (b) maintaining the solid-liquid phase admixture under biological assay conditions for a time period sufficient for the anti-apo B-100 antibodies and anti-apo A-I antibodies to immunoreact with the fusion polypeptide to form solid-phase fusion polypeptide/anti-apo B-100 and fusion polypeptide/anti-apo A-I immunocomplexes and for said antibodies to immunoreact with native apo B-100 and apo A-I, respectively, present in the sample; and
- (c) determining the amounts of solid-phase fusion polypeptide/anti-apo B-100 and fusion polypeptide/anti-apo A-I immunocomplexes formed, and thereby the amounts of native apo B-100 and native apo A-I in the sample.

48. The method of claim 47, wherein the anti-apo B-100 antibodies are MB47 antibodies conjugated to a first marker and the anti-apo A-I antibodies are AI-11 antibodies conjugated to a second marker.

49. The method of claim 47, wherein the body fluid sample, fusion polypeptide, anti-apo B-100 antibodies, and anti-apo A-I antibodies are admixed substantially simultaneously.

50. A method for assaying the amounts of native apo B-100 and native apo A-I in a body fluid sample comprising:

5 (a) admixing a body fluid sample in a liquid medium with predetermined amounts of pan anti-apo B-100 antibodies, pan anti-apo A-I antibodies, and a fusion polypeptide to form an admixture, said pan anti-apo B-100 antibodies immunoreacting with any native apo B-100 in the sample and with the fusion  
10 polypeptide, said pan anti-apo A-I antibodies immunoreacting with any native apo A-I in the sample and with the fusion polypeptide, said fusion polypeptide containing:

15 (i) a first amino acid residue sequence up through about 250 residues in length that includes the amino acid residue sequence of apo A-I shown in SEQ ID NO:3 from about residue 120 through about residue 135, said second amino acid residue  
20 sequence of said fusion polypeptide immunoreacting with anti-apo A-I antibodies secreted by a hybridoma selected from the group consisting of:

AI-4 (ATCC HB 8744),  
AI-7 (ATCC HB 8745),  
AI-9 (ATCC HB 8741),  
25 AI-10 (ATCC HB 9200),  
AI-11 (ATCC HB 9201),  
AI-12 (ATCC HB 9202),  
AI-13 (ATCC HB 9203),  
AI-14 (ATCC HB 9204), and  
30 AI-18 (ATCC HB 9570);

(ii) a second amino acid residue sequence up through about 375 residues in length that

includes the amino acid residue sequence of the apo B-100 sequence shown in SEQ ID NO:1 from about residue 217 through about residue 297, said fusion polypeptide immunoreacting with antibodies secreted by the  
5 hybridoma MB47 (ATCC HB 8746);

said second amino acid residue sequence operatively linked to the carboxy-terminus of said first amino acid residue sequence;

(b) maintaining the admixture under  
10 biological assay conditions for a time period sufficient for the anti-apo B-100 antibodies to immunoreact with the fusion polypeptide to form a fusion polypeptide/anti-apo B-100 immunocomplex and for the anti-apo A-I antibodies to immunoreact with  
15 the fusion polypeptide to form a fusion polypeptide/anti-apo A-I immunocomplex and for said antibody to immunoreact with apo B-100 and apo A-I, respectively, present in the sample; and

(c) determining the amounts of fusion  
20 polypeptide/anti-apo B-100 immunocomplex formed and the amounts of fusion polypeptide/anti-apo A-I immunocomplex formed, and thereby the amounts of native apo B-100 and native apo A-I in the body fluid sample.

25

51. The method of claim 50, wherein the anti-apo B-100 antibodies and anti-apo A-I antibodies are affixed to a solid support prior to admixing the fusion polypeptide and antibody fluid samples.

30

52. The method of claim 50, wherein the anti-apo B-100 antibodies are affixed to a first solid



support and the anti-apo A-I antibodies are affixed to a second solid support.

53. The method of claim 50, wherein the body  
5 fluid sample, fusion polypeptide, anti-apo B-100  
antibodies, and anti-apo A-I antibodies are admixed  
substantially simultaneously.

54. The method of claim 50, wherein the fusion  
10 polypeptide is conjugated to a label selected from the  
group consisting of radioactive, enzyme, biotin,  
fluorescent, and chemiluminescent labels.

GAGTCCCCAGGACCTTTCAAATTCCCTGGATACACTGTTCCAGTTGTCAATGTTGAAGTG  
GluLeuProArgThrPheGlnIleProGlyTyrThrValProValValAsnValGluVal  
aa 3214 (9642 bp) 3233 (9699)

TCTCCATTCCATAGAGATGTCGGCATTCGGCTATGTGTTCCTCCAAAGCAGTCAGCATG  
SerProPheThrIleGluMetSerAlaPheGlyTyrValPheProLysAlaValSerMet  
3253 (9759)

CCTAGTTTCTCCATCCTAGGTTCTGACGTCCGTGCTGCCCTTCATACACATTAACTCCGCCA  
ProSerPheSerIleLeuGlySerAspValArgValProSerTyrThrLeuIleLeuPro  
3273 (9819)

TCATTAGAGCTGCCAGTCCTTCATGTCCCTAGAAATCTCAAGCTTCTCTCCAGATTTC  
SerLeuGluLeuProValLeuHisValProArgAsnLeuLysLeuSerLeuProAspPhe  
3293 (9879)

AAGGAATTGTGTACCATAAGCCATATTTTATTCTCTGCCATGGGCAATATTACCTATGAT  
LysGluLeuCysThrIleSerHisIlePheIleProAlaMetGlyAsnIleThrTyrAsp  
3313 (9939)

TTCTCCTTTAAATCAAGTGTCAATCACACTGAATACCAATGCTGAACCTTTTAACCAGTCA  
PheSerPheLysSerSerValIleThrLeuAsnThrAsnAlaGluLeuPheAsnGlnSer  
3333 (9999)

FIGURE 1A

37-->  
 GATATTGTTGCTCATCTCCTTTCTTCATCTGTCTCATTTGATGCACAGCTGCAGTACAAA  
 AspIleValAlaHisLeuLeuSerSerSerValIleAspAlaLeuGlnTyrLys  
 3353 (10,059)

TTAGAGGGCACCAAGATTGACAAGAAAAGGGGATTGAAGTTAGCCACAGCTCTGTCT  
 LeuGluGlyThrThrArgLeuThrArgLysArgGlyLeuLysLeuAlaThrAlaLeuSer  
 3373 (10,119)

34-->  
 CTGAGCAACAAATTGTGGAGGGTAGTCATAACAGTACTGTGAGCTTAACACGAAAAAT  
 LeuSerAsnLysPheValGluGlySerHisAsnSerThrValSerLeuThrLysAsn  
 3393 (10,179)

ATGGAAGTGTGAGTGGCAACACCAAAAGCCCAAAATTCCTTGTGAGAAATGAATTC  
 MetGluValSerValAlaThrThrThrLysAlaGlnIleProIleLeuArgMetAsnPhe  
 3413 (10,239)

33--> 46-->  
 AAGCAAGAACTTAATGGAATACCAAGTCAAACCTACTGTCTCTTCCTCCATGGAATTT  
 LysGlnGluLeuAsnGlyAsnThrLysSerLysProThrValSerSerSerMetGluPhe  
 3433 (10,299)

FIGURE 1B

110-->  
 AAGTATGATTCAATTCTTCAATGCTGTA~~CTCTACCGCTAAAGGAGCAGTTGACCACAAG~~  
 LysTyrAspPheAsnSerSerMetLeuTyrSerThrAlaLysGlyAlaValAspHisLys  
 3453 (10,359)

111-->  
 CTTAGCTTGGAAAGCCTCACCTCTTACTTTTCCATTGAGTCATCTACCAAAGGAGATGTC  
 LeuSerLeuGluSerLeuThrSerTyrPheSerIleGluSerSerThrLysGlyAspVal  
 3473 (10,419)

113-->  
 AAGGGTTCGGTCTTTTCTCGGGAATATTCAGGA~~ACTATTGCTAGTGAGGCCAACACTTAC~~  
 LysGlySerValLeuSerArgGluTyrSerGlyThrIleAlaSerGluAlaAsnThrTyr  
 3493 (10,479)

114-->  
 99<--  
 TTGAA~~TTCC~~AAGAGCACACGGTCTTCAGTGAAGCTGCAGGGC~~ACTTCCAA~~AAATTGATGAT  
 LeuAsnSerLysSerThrArgSerSerValLysLeuGlnGlyThrSerLysIleAspAsp  
 3513 (10,539)

135<--  
 ATCTGGAACCTTGAAGTAAAGAAAATTTTGCTGGAGAGCCACACTCCAACGCATATAT  
 IleTrpAsnLeuGluValLysGluAsnPheAlaGlyGluAlaThrLeuGlnArgIleTyr  
 3533 (10,599)

FIGURE 1C

134<--  
TCCCTCTGGGAGCACAGTACGAAACCACTTACAGCTAGAGGGCCTCTTTTCACCAAC  
SerLeuTrpGluHisSerThrLysAsnHisLeuGlnLeuGluGlyLeuPhePheThrAsn  
3553 (10,659)

133<--  
GGAGAACATACAAGCAAAGCCACCCCTGGAACTCTCTCCATGGCAAATGTCAGCTCTTGTT  
GlyGluHisThrSerLysAlaThrLeuGluLeuSerProTrpGlnMetSerAlaLeuVal  
3573 (10,719)

130<--  
CAGGTCCATGCAAGTCAGCCAGTTCCTTCCATGATTCCCTGACCTTGGC  
GlnValHisAlaSerGlnProSerSerPheHisAspPheProAspLeuGly  
3590 (10,770p)

FIGURE 1D

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ATG AAA GCT GCG GTG CTG ACC TTG GCC GTG CTC TTC CTG ACG GGG AGC	48
Met Lys Ala Ala Val Leu Thr Leu Ala Val Leu Phe Leu Thr Gly Ser	15
	10
	5
CAG GCT CGG CAT TTC TGG CAG CAA GAT GAA CCC CCC CAG AGC CCC TGG	96
Gln Ala Arg His Phe Trp Gln Gln Asp Gln Pro Pro Gln Ser Pro Trp	30
	25
	20
GAT CGA GTG AAG GAC CTG GCC ACT GTG TAC GTG GAT GTG CTC AAA GAC	144
Asp Arg Val Lys Asp Leu Ala Thr Val Tyr Val Asp Val Leu Lys Asp	45
	40
	35
AGC GGC AGA GAC TAT GTG TCC CAG TTT GAA GGC TCC GCC TTG GGA AAA	192
Ser Gly Arg Asp Tyr Val Ser Gln Phe Glu Gly Ser Ala Leu Gly Lys	60
	55
	50
CAG CTA AAC CTA AAG CTC CTT GAC AAC TGG GAC AGC GTG ACC TCC ACC	240
Gln Leu Asn Leu Lys Leu Leu Asp Asn Trp Asp Ser Val Thr Ser Thr	80
	75
	70
	65
TTC AGC AAG CTG CGC GAA CAG CTC GGC CCT GTG ACC CAG GAG TTC TGG	288
Phe Ser Lys Leu Arg Glu Gln Leu Gly Pro Val Thr Gln Glu Phe Trp	95
	90
	85

FIGURE 2A

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GAT AAC CTG GAA AAG GAG ACA GAG GGC CTG AGG CAG GAG ATG AGC AAG	336
Asp Asn Leu Glu Lys Glu Thr Glu Gly Leu Arg Gln Glu Met Ser Lys	110
	105
	100
GAT CTG GAG GAG GTG AAG GCC AAG GTG CAG CCC TAC CTG GAC GAC TTC	384
Asp Leu Glu Glu Val Lys Ala Lys Val Gln Pro Tyr Leu Asp Asp Phe	125
	115
	120
CAG AAG AAG TGG CAG GAG GAG ATG GAG CTC TAC CGC CAG AAG GTG GAG	432
Gln Lys Lys Trp Gln Glu Glu Met Glu Leu Tyr Arg Gln Lys Val Glu	140
	135
	130
CCG CTG CGC GCA GAG CTG CAA GAG GGC GCG CAG AAG CTG CAC GAG	480
Pro Leu Arg Ala Glu Leu Gln Glu Gly Ala Arg Gln Lys Leu His Glu	155
	150
	145
CTG CAA GAG AAG CTG AGC CCA CTG GGC GAG GAG ATG CGC GAC CGC GCG	528
Leu Gln Glu Lys Leu Ser Pro Leu Leu Gly Glu Glu Met Arg Asp Arg Ala	175
	165
	170
CGC GCC CAT GTG GAC GCG CTG CGC ACG ACG CAT CTG GCC CCC TAC AGC GAC	576
Arg Ala His Val Asp Ala Leu Arg Thr His Leu Ala Pro Tyr Ser Asp	190
	185
	180

FIGURE 2B

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GAG CTG	CGC	CAG	CGC	TTG	GCC	GCG	CGC	CTT	GAG	GCT	CTC	AAG	GAG	AAC	624
Glu Leu	Arg	Gln	Arg	Leu	Ala	Ala	Arg	Leu	Glu	Ala	Leu	Lys	Glu	Asn	
	195					200					205				
GGC GGC	GCC	AGA	CTG	GCC	GAG	TAC	CAC	GCC	AAG	GCC	ACC	GAG	CAT	CTG	672
Gly Gly	Ala	Arg	Leu	Ala	Glu	Tyr	His	Ala	Lys	Ala	Thr	Glu	His	Leu	
	210				215					220					
AGC ACG	CTC	AGC	GAG	AAG	GCC	AAG	CCC	GCG	CTC	GAG	GAC	CTC	CGC	CAA	720
Ser Thr	Leu	Ser	Glu	Lys	Ala	Lys	Pro	Ala	Leu	Glu	Asp	Leu	Arg	Gln	
	225			230					235					240	
GGC CTG	CTG	CCC	GTG	CTG	GAG	AGC	AGC	TTC	AAG	GTC	AGC	TTC	CTG	AGC	GCT 768
Gly Leu	Leu	Pro	Val	Leu	Glu	Ser	Phe	Lys	Val	Ser	Phe	Leu	Ser	Ala	
			245					250					255		
CTC GAG	GAG	TAC	ACT	AAG	AAG	CTC	AAC	ACC	CAG						801
Leu Glu	Glu	Tyr	Thr	Lys	Lys	Leu	Asn	Thr	Gln						
		260					265								

FIGURE 2C



## INTERNATIONAL SEARCH REPORT

International application No.  
PCT/US92/08634

**A. CLASSIFICATION OF SUBJECT MATTER**

IPC(5) :C07H 21/02; C07K 15/06; C12N 15/70; G01N 33/53

US CL :530/350, 359; 536/27; 435/7.1, 320.1

According to International Patent Classification (IPC) or to both national classification and IPC

**B. FIELDS SEARCHED**

Minimum documentation searched (classification system followed by classification symbols)

U.S. : 530/350, 359; 536/27; 435/7.1, 320.1

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practicable, search terms used)

MEDLINE, CAS, BIOSIS, APS,

search terms: apo B-100, apo A-1, expression, recombinant DNA, E. coli, human, apolipoprotein

**C. DOCUMENTS CONSIDERED TO BE RELEVANT**

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	DNA, Volume 8, No.6, issued 1989, N. Moguilevsky et al, "Production of Human Recombinant Proapolipoprotein A-I in <u>Escherichia coli</u> : Purification and Biochemical Characterization", pages 429-436, especially Figure 2.	15-18
Y	The EMBO Journal, Volume 6, No. 11, issued 1987, L. Monaco et al, "A recombinant apoA-I-protein A hybrid reproduces the binding parameters of HDL to its receptor", pages 3253-3260, especially Figure 1 and Table II.	19-28, 31-54
<del>X</del> Y	The Journal of Biological Chemistry, Volume 265, No. 1, issued 05 January 1990, R. J. Pease et al., "Use of Bacterial Expression Cloning to Localize the Epitopes for a Series of Monoclonal Antibodies against Apolipoprotein B-100", pages 553-568, see entire document.	<u>10-13</u> 1-9, 14, 22-28, 31-54
Y	Clinical Chemistry, Volume 32, No. 8, issue 1986, S. G. Young et al, "Two New Monoclonal Antibody-Based Enzyme-Linked Assays of Apolipoprotein B", pages 1484-1490, see entire document.	27, 28, 31-54

☒ Further documents are listed in the continuation of Box C.

☐ See patent family annex.

Special categories of cited documents:	
*A* document defining the general state of the art which is not considered to be part of particular relevance	*T* later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention
*E* earlier document published on or after the international filing date	*X* document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone
*L* document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)	*Y* document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art
*O* document referring to an oral disclosure, use, exhibition or other means	
*P* document published prior to the international filing date but later than the priority date claimed	*Z* document member of the same patent family

Date of the actual completion of the international search

18 DECEMBER 1992

Date of mailing of the international search report

12 JAN 1993

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## INTERNATIONAL SEARCH REPORT

International application No.  
PCT/US92/08634

## C (Continuation). DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
Y	EP, A, 0,301,667 (BARAELE ET AL) 01 February 1989, pages 2-5, 7-11, and Figures 3-7.	27, 28, 31-54
Y	WO, A, 86/06742 (SCHENK ET AL) 20 November 1986, pages 6-9, Example II, VI, VII, and Figures 1 and 2.	38-54

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